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(57) Abstract  The present invention concerns essential genes from	ı C. albi	cans and their use in a method for the screening of antimycotic substances.

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# ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCREENING ANTIMYCOTIC SUBSTANCES USING SAID GENES

The present invention relates to a method for screening for antimycotic substances in which essential genes from mycetes, particularly from Candida albicans (C.albicans) as well as functionally similar genes from other pathogenic mycetes, or the corresponding encoded proteins, are used as targets. The invention also relates to specific C. albicans genes.

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The spectrum of known fungal infections stretches from fungal attack of skin or nails to potentially hazardous mycotic infections of the inner organs; Such infections and resulting diseases are known as mycosis.

Antimycotic substances (fungistatic or fungicidal) are 15 used for treatment of mycosis. However, up to now, relatively few substances with pharmacological effects are Nystatin, Pimaricin, Amphotericin B, such as known. 5-fluoro-cytosine and Griseofulvin, Clotrimazole, Batraphene. The drug treatment of fungal infections is 20 extremely difficult, in particular because both the host cells and the mycetes, are eucaryotic cells. Administration of drugs based on known antimycotic substances results therefore often in undesired side-effects, for example Amphotericin B has a nephrotoxic effect. Therefore, there 25 is a strong need for pharmacologically efficient substances usable for the preparation of drugs, which are suitable for prophylactic treatments of immunodepressive states or for the treatment of an existing fungal infection. Furthermore, the substances should exhibit a specific spectrum of action 30 selectively inhibit the growth and order to proliferation of mycetes without affecting the treated host organism.

The aim of the present invention is to provide a method for the identification of antimycotic substances and especially for the identification of anti-Candida substances. An essential feature of this method is that essential genes from mycetes are used as targets for the screening.

The present invention thus concerns a method for screening antimycotic substances wherein an essential gene from mycetes or a functionally similar gene in another pathogenic mycete, or the corresponding encoded protein, is used as target and wherein the essential gene is selected from the group consisting of CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 and CaJL039.

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According to one embodiment of the method of the invention mycete cells which express the essential gene, or a functionally similar mycete gene, to a different level are incubated with the substance to be tested and the growth inhibiting effect of the substance is determined.

According to another embodiment, said target gene or the corresponding target gene encoded protein is contacted in vitro with the substance to be tested and the effect of the substance on the target is determined.

According to another embodiment, the screened substances inhibit partially or totally the functional expression of the essential genes or the functional activity of the encoded proteins.

According to one embodiment the screened substances partially or totally inhibit the activity of dihydropteroate synthase (DHPS) and/or 7,8-dihydro-6-hydroxymethylpterin-pyrophosphokinase (HPPK).

According to another embodiment, the mycete species are selected from the group comprising Basidiomycetes, Ascomycetes and Hyphomycetes.

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According to another embodiment of the method of the invention said functional similar genes are essential genes from Candida Spp., preferably Candida albicans, or from Aspergillus Spp., preferably from Aspergillus fumigatus.

According to another aspect, the present invention concerns a polynucleotide having the sequence as depicted in SEQ ID No.2, SEQ ID No.4, SEQ ID No.6, SEQ ID No.7, SEQ ID No.9, SEQ ID No.10, SEQ ID No.11 or SEQ ID No.13, preferably SEQ ID No.2, SEQ ID No.4, SEQ ID No.6, SEQ ID No.9, SEQ ID No.10 or SEQ ID No.11, homologs thereof and functional fragments thereof.

According to another aspect, the present invention concerns a gene which is CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, preferably CaOR110, CaMR212, CaNL256, CaBR102 or CaIR012, or a functionally similar gene or a functional fragment thereof.

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According to this embodiment, the functionally similar gene or homologous polynucleotide has a sequence identity, at the nucleotide level, with CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, respectively, of at least 50%, preferably of at least 60%, and most preferably of at least 70%. A functional fragment is a polynucleotide fragment that will retain the functionality of the starting product (nucleotide or gene). One example is the CaOR110 splice variant (which is also homologous to the original gene, with about 90% identity).

According to another embodiment, the functionally similar gene has a sequence identity, at the amino-acid level, with CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, respectively, encoded protein(s) of at least 40%, preferably of at least 50%, more preferably of at least 60% and most preferably of at least 70%.

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These figures given for the gene apply mutatis mutandis to the polynucleotide, as far as homology and similarity.

According to another aspect, the present invention covers the protein(s) encoded by CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, respectively, gene(s) or by a functionally similar gene, or a functional polypeptidic fragment thereof.

According to another aspect, the present invention provides a plasmid containing CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, respectively, gene(s), a functionally similar gene or a functional fragment thereof

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According to another aspect, the present invention provides a plasmid (bacteria containing same) deposited at the CNCM (Institut Pasteur, Paris) on 98/08/13, with the accession numbers I-2065, I-2063 and I-2064, corresponding to the CaNL256, CaBR102 and CaIR012 genes, respectively.

According to another aspect, the present invention provides a plasmid (bacteria containing same) deposited at the DSMZ (Deutsche Sammlung von Mikroorganismen und Zellkulturen, Germany) on 99/08/06 with the accession numbers DSM 12977, DSM 12976, DSM 12978 and DSM 12979, corresponding to the CaDR325, CaOR110, CaOR110 splice variant and CaMR212, respectively.

According to another aspect, the present invention provides a kit for diagnosis of fungal infections comprising a gene selected from the group consisting of CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 and CaJL039, a functionally similar gene thereof, a functional fragment thereof, the corresponding encoded protein or a functional polypeptide fragment thereof.

According to another aspect, the present invention provides an antibody directed against the protein encoded

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by the CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, respectively, gene(s) or by a functionally similar gene, or a polypeptide fragment thereof.

According to another aspect, the present invention provides a polynucleotide obtainable by the process comprising the following steps:

- (i) selecting an essential gene from Saccharomyces cerevisiae;
- (ii) comparing the sequence of said gene with
  10 Candida Albicans genome sequences;
  - (iii) deducing homologuous oligonucleotides
    regions;
  - (iv) PCR amplifying the thus-obtained
    oligonucleotides;
- 15 (v) using the amplimers of step (iv) for detecting the complete gene of interest:

the amplimers of step (iv) are used as a probe for detecting the complete gene of interest from a Candida albicans genomic or cDNA library; or

20 the complete gene is obtained by 3' and 5' extension of the amplimer, e.g. by using a PCR method.

According to the invention, the first step is to identify said essential genes and starting from these thus identified genes, essential genes from other pathogenic mycetes can be identified. For practical purposes, essential genes from S. cerevisiae are first identified and starting from them, essential genes from other pathogenic fungus, especially from Candida, are obtained.

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The present invention thus discloses the identification of essential genes from C.albicans and their use in a method for the screening of antimycotic substances, especially anti-Candida substances.

In order to identify essential genes of S.cerevisiae, individual genomic genes are eliminated through homologous

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recombination. If the DNA segment thus eliminated concerns an essential gene, then the deletion is lethal for the S.cerevisiae cells in haploid form.

A method, wherein the studied S. cerevisiae gene is replaced by a marker gene can be used to generate the corresponding genomic deletion of S.cerevisiae and to determine the S.cerevisiae cells containing the deletion.

As a selection marker a dominant selection marker (e.g. kanamycin resistance gene) or an auxotrophic marker can for example be used. As an auxotrophic marker, it is possible to use genes coding for key enzymes of amino acid or nucleic base synthesis. For example, one can use as a selection marker the following genes from S.cerevisiae: gene encoding for the metabolic pathway of leucine (e.g.LEU2-gene), histidine (e.g. HIS3-gene) or tryptophan (e.g. TRP-1 gene) or for the nucleic base metabolism of uracil (e.g. URA3-gene).

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Auxotrophic S.cerevisiae strains can be used. These auxotrophic strains can only grow on nutritive media containing the corresponding amino acids or nucleotide bases. All laboratory S.cerevisiae strains, containing auxotrophic markers can for instance be used. When diploid S.cerevisiae strains are used, then the corresponding marker gene must be homozygously mutated. Strain CEN.PK2 or isogenic derivates thereof can be used.

Strains containing no suitable auxotrophic marker can also be used such as prototrophic S.cerevisiae strains. Then a dominant selection marker e.g. resistance gene, such as kanamycin resistance gene can be used. A loxP-KanMX-loxP cassette can advantageously be used for this purpose.

For the homologous recombination replacing the whole DNA sequence or part thereof of a S.cerevisiae gene, DNA fragments are used wherein the marker gene is flanked at

the 5'- and 3'-ends by sequences which are homologous to the 5'- and 3'-ends of the studied S.cerevisiae gene.

Different processes can be used for the preparation of the corresponding DNA fragments which are also appropriate for the deletion of any specific S.cerevisiae gene. A linear DNA-fragment is used for the transformation of the suitable S.cerevisiae strain. This fragment is integrated into the S.cerevisiae genome by homologous recombination. These processes include:

- 10 1. "Conventional method" for the preparation of deletion cassettes (Rothstein, R.J. (1983) Methods in Enzymology, Vol. 101, 202-211).
  - 2. "Conventional Method" using the PCR technique ("modified conventional method").
- 3. SFH (short flanking homology) PCR method (Wach, A. et al. (1994) Yeast 10: 1793-1808; Gültner, U. et al. (1996) Nucleic Acids Research 24:2519-2524).
  - 1. In the "conventional method" for the preparation of deletion cassettes in the S.cerevisiae genome, the gene to be studied is either already present in an appropriate vector or is integrated in such a vector. With this method, any pBR- pUC- and pBluescript®-derivates can be used for example. A major part of the target gene sequence is eliminated from the vector, for instance using appropriate restriction sites, conserving however the 3'- and 5'-regions of the studied gene inside the vector. The selected marker gene is integrated between the remaining regions.
  - 2. In the modified form of this "conventional method", PCR is used. This method allows amplification of the 3'-and 5'-terminal regions of the coding sequence of the studied S.cerevisiae gene. This method amplifies selectively both terminal regions of the studied gene, therefore, two PCR-reactions must be carried out for each studied gene, amplifying once the 5'-end, and once the 3'-

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end of the gene. The length of the amplified terminal DNA-fragment depends on the existing restriction sites. The amplified terminal ends of the studied gene have generally a length of 50 to 5000 base pairs (bp), preferably a length between 500 and 1000 bp.

As template for the PCR-reactions, genomic DNA of S.cerevisiae or wild-type genes can be used. The primer-pairs (a sense and an antisense primer, respectively) are constructed so that they correspond to the 3'-end and the 5'-end sequence of the studied S.cerevisiae gene. Especially, the primer is selected such as to allow its integration by way of appropriate restriction sites.

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As vectors, pBR- pUC- and pBluescript®-derivates can be used. In particular vectors already containing a gene encoding the selection marker, are appropriate. In particular, vectors can be used, which contain genes of the selection marker HIS3, LEU2, TRP1 or URA3.

The DNA segments of the studied S.cerevisiae gene, obtained by PCR, are integrated in the vector at both sides of the selection marker, so that subsequently, as in the "conventional method", the selection marker is flanked on both ends by DNA sequences which are homologous to the studied gene.

place in a very efficient and precise manner and the length of the DNA sequence homologous to the studied S.cerevisiae gene flanking the selection marker gene can in fact be considerably shorter than with the "modified conventional method". The flanking ends homologous to the studied S.cerevisiae gene need to present a length of only about 20-60 bp, preferably 30-45 bp. The SFH-PCR method is particularly advantageous as the laborious cloning step can be obviated.

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A PCR reaction is carried out on a DNA-template containing the gene for the selection marker to be used, wherein the primers are constructed so that the DNA sequence of the sense primer is homologous to the 5'-end of the selection marker sequence and so that the primer presents in addition at its 5'-end a region of preferably which corresponds to the 5'-terminal 40 nucleotides, sequence of the studied S.cerevisiae gene. The antisense primer is constructed in an analogous manner, i.e. it is complementary to the 3'-end of the gene sequence of the selection marker, wherein this primer contains at its 5'end a region of also preferably 40 nucleotides, which corresponds to the complementary strand of the 3'-terminal coding sequence of the studied gene.

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For the amplification of S.cerevisiae genes to be studied by the SFH-PCR method, vectors containing the gene for the auxotrophic marker or selection marker can be used. Especially, plasmid pUG6 is used as the template. This plasmid contains a loxP-KanMX-loxP cassette (Gültner, U. et al. (1996) Nucleic Acids Research 24: 2519-2524). In other terms, the Kanamycin resistance gene is flanked at both ends by a loxP sequence (loxP-KanMX-loxP cassette). cassette is advantageous in that the Kanamycin resistance gene can be eventually eliminated from the S.cerevisiae genome after integration of the loxP-KanMX-loxP cassette into the S.cerevisiae gene to be studied. Cre-recombinase of bacteriophage P1 can be used for this purpose. Crerecombinase recognizes the loxP sequences and induces elimination of the DNA located between the two loxP sequences by a homologous recombination process. As a result only one loxP sequence remains and the so-called marker regeneration occurs, i.e. the S.cerevisiae strain again using the loxP-KanMX-loxP transformed cassette. This is particularly advantageous, when at least

two functionally similar genes are to be deleted in order to obtain a lethal phenotype.

With the PCR-method, the PCR reaction primers are at the 3'-end a preferably 20 nucleotide long sequence, which is homologous to the sequence situated left and/or right of at the loxP-KanMX-loxP cassette, and preferably 40 nucleotide long sequence, which is homologous to the terminal ends of the gene to be studied.

Using the three methods, one obtains linear deletion cassettes containing the gene encoding the selection marker, which is flanked on both sides by homologous sequences of the gene to be studied. The deletion cassettes are used for the transformation of diploid S.cerevisiae S.cerevisiae strain diploid The strains. (Scientific Research & Development GmbH, Oberursel) can be used for example for this purpose.

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[CEN.PK2 Mata/MAT  $\alpha$  ura3-52/ura3-52 leu2-3, 112/leu2-112his3Δ1/his3Δ1 trp1-289/trp1-289 MAL2-8<sup>C</sup>/MAL2-8<sup>C</sup> 3. SUC2/SUC2]

The strain CEN.PK2 is prepared and cultivated using 20 known methods (Gietz, R.D. et al. (1992) Nucleic Acids Research 8: 1425; Güldener, U. et al. (1996) Nucleic Acids Research 24:2519-2524).

The cells of the S.cerevisiae strain used are with according to processes known 25 ±ransformed appropriate DNA quantity of the linear deletion cassette (e.g. Sambrook et al. 1989). Thereafter, the medium in which the cells are cultivated is replaced by a new medium, a so-called selective medium, which does not contain the corresponding amino acid (e. g. histidine, leucine or tryptophan) or nucleic base (e. g. uracil) or, when using a deletion cassette containing the kanamycin resistance gene, by a medium containing geneticin (G418 $^{\odot}$ ) (e.g. a complete medium (YEPD) containing geneticin). Alternatively, the 10

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transformed cells may be plated on agar plates prepared using the corresponding media. Thereby, one selects the transformed cells, in which a homologous recombination occured, since only those cells can grow under these modified conditions.

However, in most cases, only one of the two copies of the gene in the double chromosome set of a diploid S.cerevisiae strain is replaced by the DNA of the deletion cassette during the transformation, resulting heterozygote-diploid S.cerevisiae mutant strain, wherein one copy of the gene studied is replaced by a selection marker, while the other copy of the gene is maintained in the genome. This presents the advantage that in case of a deletion of an essential gene, due to the existence of the second copy of the essential gene, the mutant S.cerevisiae strain is still viable.

The proper integration of the deletion cassette DNA at the predetermined chromosomal gene locus (gene locus of the gene to be studied) may be checked by Southern-Blot Analysis (Southern, E.M. (1975) J. Mol. Biol. 98:503-517) or by diagnostic PCR analysis using specific primers (Güldener, U. et al. (1996) Nucleic Acids Research 24:2519-2524)

The genetic separation of individual diploid cells may be monitored by tetrad analysis. To this end, reduction division (meiosis) is induced in the diploid cells, especially heterozygote mutant strains, using known methods such as nitrogen impoverishment on potassium acetate plates (Sherman, F. et al. (1986) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y.; Guthrie, C. and Fink, G.R. (1991) Methods in Enzymology, Vol 194. Academic Press, San Diego, 3-21; Ausubel, F. M. et al. (1987) Current Protocol in Molecular Biology John Wiley and Sons, Inc., Chapter 13). Meiosis results only in asci with four 15

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ascospores (segregated), which can be indivualized after partial enzymatic digestion of the ascospore wall with way (1987)) by al. (Ausubel et zymolyase micromanipulators (e.g. SINGER). For example when a tetrad analysis is carried out on a heterozygote-diploid mutant strain in which an essential gene present in the double chromosome set is replaced on one chromosome by homologous recombination, then only two segregated ascospores are viable, namely those which carry the essential gene. The 10 two remaining segregated ascospores are not viable because they lack the essential gene.

In order to check if the genes studied by this method are really essential or if the homologous recombination leads to an alteration of an essential gene adjacent to the gene locus of the gene studied, the heterozygote diploid S.cerevisiae mutant strain is transformed with a centromere plasmid containing said studied gene.

A tetrad analysis is carried out on the transformants. When four instead of two viable segregates are obtained, then the studied gene contained in the centromere plasmid can complement the defect of the two non-viable haploid S.cerevisiae cells/mutant strains, which demonstrates that the studied S.cerevisiae gene is essential.

Preferably, plasmids present in low copy number, e.g. one or two copies per cell are used as centromere plasmids. For example plasmids pRS313, pRS314, pRS315 and pRS316 (Sijkorski, R. S. and Hieter, P. (1989) Genetics 122: 19-27) or similar plasmids can be used for this purpose. Preferably, the studied genes are integrated in said plasmids including their 3'- and 5'-end non-coding regions.

Individual S.cerevisiae genes may be studied using the above-described method, their sequences being totally or partially known. The complete genomic sequence of

S.cerevisiae was made accessible to the public via the WWW (World Wide Web) on April 24, 1996.

Different possibilities exist to have access to the S.cerevisiae genomic DNA sequence via the WWW.

MIPS (Munich information Centre of Protein Sequence)
Address <a href="http://speedy.mips.biochem.mpg.de/mips">http://speedy.mips.biochem.mpg.de/mips</a> /yeast/

SGD (Saccharomyces Genome Database, Stanford)

Address http://genome-www.stanford.edu/Saccharomyces

YPD(Yeast Protein Database, Cold Spring Harbor)

Address http://www.proteome.com/YPDhome.html

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The complete S.cerevisiae DNA sequence is also accessible via FTP (file transfer protocol) in Europe (e.g. at the address: ftp.mips.embnet.org) in the U.S.A. (address: genome-ftp.stanford.edu) or in Japan (address: ftp.nig.ac.jp).

7 essential genomic S.cerevisiae genes have been identified by this way: YDR325w, YJL039c, YOR110w, YNL256w, YBR102c, YIR012w and YMR212c

The essential genes of S.cerevisiae are then used to identify corresponding functionally similar genes in other mycetes.

By functionally similar genes in other mycete species, is meant genes which have a function similar or identical to that of the identified essential genes of S.cerevisiae. Functionally similar genes in other mycetes may, but need not be homologous in sequence to the corresponding essential S.cerevisiae genes. Functionally similar genes in other mycetes may exhibit only moderate sequence homology at the nucleotide level to the corresponding essential S.cerevisiae genes. By moderate sequence homology it is meant in the present invention genes having a sequence identity, at the nucleotide level, of at least 50%, more preferably of at least 60% and most preferably of at least 70%.

In addition, functionally similar genes in other mycetes may, but need not encode proteins homologous in sequence to the proteins encoded by the essential S.cerevisiae genes. Functionally similar proteins in other mycetes may exhibit moderate protein sequence homology to the proteins encoded by the essential S.cerevisiae genes.

By moderate protein sequence homology is meant in the present invention proteins having a sequence identity, at the amino-acid level, of a least 40%, preferably of at least 50%, more preferably of at least 60% and most preferably of at least 70%.

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Genes homologous in sequence may be isolated according to known methods, for example via homologous screening (Sambrook, J. et al. (1989) Molecular Cloning Cold Spring Harbor Laboratory Press, N.Y.) or via the PCR technique using specific primers from genomic libraries and/or cDNA libraries of the corresponding mycetes.

According to one embodiment, genes homologous in sequences are isolated from cDNA libraries. In order to find out functionally similar genes in other mycetes, mRNA is isolated from mycete species to be studied according to known methods (Sambrock et al. 1989) and cDNA is synthesized according to known methods (Sambrock et al. 1989; or cDNA synthesis kits, e.g. from STRATAGENE).

The prepared cDNA is directionally integrated in a suitable expression vector.

For example, synthesis of the first cDNA strand may be carried out in the presence of primers having appropriate restriction sites in order to allow a subsequent cloning in the proper orientation with respect to the expression vector promoter. As restriction sites, any known restriction site may be used. As a primer, for instance the following primer, 50 nucleotides long may be used:

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appropriate  $(X)_{6}$ represents an The sequence restriction site, for example for XhoI.

After two-strand synthesis, the cohesive ends of the double stranded cDNA are filled (blunt end) and the cDNA ends are then ligated using a suitable DNA adaptor The DNA adaptor sequence should contain a sequence. restriction site which should be different from the restriction site used in the primer for the synthesis of the first cDNA strand. The DNA adaptor may comprise for example complementary 9- or 13-mer oligonucleotides, whose ends represent the cohesive end of a restriction site. These ends may be for example a EcoRI-site:

XXXXXGGCACGAG 3'

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XCCGTGCTC 5' 3 '

single-stranded X the adaptor sequence in represent the cohesive end of a restriction site.

The cDNA provided with corresponding adaptor sequences is then cleaved using restriction endonuclease, whose recognition site was used in the primer for the synthesis of the first cDNA strand, for example XhoI. The cDNA thus obtained would have according to this example 3'-XhoI and therefore could be ends and protruding 5'-EcoRI directionally integrated into an expression vector cleaved with XhoI and EcoRI.

others, E. among expression vectors, coli/S.cerevisiae shuttle vectors, i.e. vectors usable in E. coli as well as in S.cerevisiae are suitable. Such vectors may then be amplified for instance in E. coli. As expression vectors, those which are present in a high copy number as well as those present in a low copy number in S.cerevisiae cells can be used. For this purpose, for example vectors selected in the group consisting of pRS423

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- pRS426 (pRS423, pRS424, pRS425, pRS426) and/or pRS313pRS316 (pRS313, pRS314, pRS315, pRS316) (Sikorki, R.S. and Hieter, P. (1989) Genetics 122: 19-27; Christianson T. W. et al. (1992) Gene 110: 119-122) are suitable.

appropriate contain should Expression vectors S.cerevisiae promoters and terminators. In case they do not have these elements, the corresponding promoters terminators are inserted in such a way that a subsequent incorporation of the generated cDNA remains possible. Particularly suitable are the promoters of S.cerevisiae genes MET25, PGK1, TPI1, TDH3, ADHI, URA3. One may use promoters of the wild-type gene in non modified form as well as promoters which were modified in such a way that certain activator sequences and/or repressor sequences were eliminated. As terminators, for example the terminators of the S.cerevisiae genes MET25, PGK1, TPI1, TDH3, ADHI, URA3 are suitable.

According to another embodiment, genes homologous in sequence are isolated from genomic libraries. Genomic DNA libraries from mycetes can be prepared according to procedures known (for example as described in Current Protocols in Molecular Biology, John Wiley and Sons, Inc). For example, genomic DNA from mycetes can be prepared using known methods for yeast cell lysis and isolation of genomic DNA (for example commercially available kits from Bio101, Inc). The genomic DNA can be partially digested using a restriction enzyme such as Sau3AI and the fragments are size-selected by agarose gel electrophoresis. DNA fragments having for example a size of 5-10kb are then purified by classical methods (as for example, using Gene Clean kit from Bio101) and inserted in a E.coli/yeast shuttle vector such as YEP24 (described e.g. by Sanglard D., Kuchler K., Bille and J-L., Monod Μ. Pagani Ischer F., Antimicrobial Agents and Chemotherapy, (1995) Vol.39 No11,

P2378-2386) cut by a restriction enzyme giving compatible ends (for example BamHI for Sau3AI-cut genomic DNA). The resulting expression library can be amplified in E.coli. However any known method, appropriate for the preparation of a genomic library, can be used in the present invention.

In order to find the genes in the studied mycete species, which are functionally similar to essential genes of S.cerevisiae, one S.cerevisiae essential gene is placed under control of a regulated promoter, either as an integrative (1) or extrachromosomal (2) gene.

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1. For the integration of a regulated promoter in the S.cerevisiae genome, one replaces the native promoter of the selected essential gene by the regulated promoter, for example by homologous recombination via PCR (Güldener et al. (1996). The homologous recombination via PCR can be carried out for example in the diploid S.cerevisiae strain CEN.PK2. The successfull integration into one chromosome can be checked in haploid cells following tetrad analysis.

Using the tetrad analysis, one obtains four viable ascospores, wherein in two haploid segregates, the selected essential gene is placed under the control of the native promoter, while the essential gene in the two remaining segregates is placed under the control of the regulated promoter.

The last mentioned haploid segregates are used for the transformation with the cDNA or the genomic DNA present in the recombinant vector.

2. Using the extrachromosomal variant, the selected essential S.cerevisiae gene is first inserted in a suitable expression vector, for example a E.coli/ S.cerevisiae shuttle vector. For this purpose, the essential gene may be amplified via PCR from genomic S.cerevisiae DNA starting from the ATG initiation codon up to and including the termination codon. The primers used for this purpose may be

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constructed in such a way that they contain recognition sites for appropriate restriction enzymes, facilitating a subsequent insertion under control of a regulated promoter in an expression vector.

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The recombinant expression vector with the plasmid copy of the essential S.cerevisiae gene under the control of a regulated promoter is subsequently used for the transcomplementation of the corresponding mutant allele. The corresponding mutant allele may be selected from the heterozygote-diploid mutant strains prepared homologous totally, by or eliminating, partially recombination an essential mycete gene listed above and as described above.

The expression vector with the selected essential S.cerevisiae gene is transformed in the corresponding 15 heterozygote-diploid mutant strain carrying instead of the selected essential S.cerevisiae gene, a selection marker gene. The transformants are isolated by selection based on the auxotrophic marker contained in the expression vector 20 used. The thus transformed heterozygote-diploid mutant strains are submitted to a tetrad analysis. One obtains four viable segregates. By retracing the corresponding markers of the mutant allele and the expression vector, the transformed wild-type segregates may be distinguished from segregates which do not contain the genomic copy of the 25 essential gene. Segregates, which do not contain the genomic copy of the selected essential gene, are designated as trans-complemented haploid mutant strains. They are subsequently used for transformation with cDNA or genomic DNA libraries from other mycete species present 30 appropriate vectors.

As regulated promoters, inducible or repressible promoters may be used. These promoters can consist of naturally and/or artificially disposed promoter sequences.

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As regulated promoters, for example the promoters of GAL1 gene and the corresponding promoter derivatives, such as for example promoters, whose different UAS (upstream activation sequence) elements have been eliminated (GALS, GALL; Mumberg, J. et al. (1994) Nucleic Acids Research As regulated promoters, 22:5767-5768) may be used. promoters of gluconeogenic genes may also be used, such as e.g. FBP1, PCK1, ICL1 or parts therefrom, such as e.g. their activation sequence (UAS1 and/or UAS2) or repression sequence (URS, upstream repression sequence) (Niederacher et al. (1992), Curr. Genet. 22: 636-670; Proft et al. (1995) Mol. Gen. Gent. 246: 367-373; Schüller et al. (1992) EMBO J; 11: 107-114; Guarente et al. (1984) Cell 36: 503-511).

A S.cerevisiae mutant strain modified in this manner can be cultivated under growth conditions, in which the regulated promoter is active, so that the essential S.cerevisiae gene is expressed. The S.cerevisiae cells are then transformed with a representative quantity of the library containing the studied mycete species cDNA or genomic DNA. Transformants express additionally the protein whose coding sequence is present in the recombinant vector.

The method contemplates that the growth conditions may be modified in such a way as to inhibit the regulated promoter, under the control of which is the selected essential gene. Especially, growth conditions may be changed by replacing the growth medium. When for example the GAL1 promoter or a derivate thereof is used, one can replace the galactose-containing medium (induced state) by a glucose-containing medium (repressed state).

These modified conditions are lethal for the S.cerevisiae cells in which the recombinant vector does not carry the functionally similar genomic DNA or cDNA of the studied mycete species. On the contrary, the S.cerevisiae

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recombinant vector expresses in which the cells functionally similar coding sequence of the studied mycete species, are viable, since in these cells the lethal metabolic defect is complemented by the protein encoded by the functionally similar gene.

The method contemplates that the recombinant vector (the plasmid) is isolated from the surviving transformants using known method (Strathern, J.N. and Higgins, (1991). Plasmids are recovered from yeast into Escherichia coli shuttle vectors in: Guthrie, C. and Fink, G.R. Methods in Enzymology, Volume 194. Guide to yeast genetic and molecular Biology. Academic Press, San Diego, 319-329) and the cDNA or genomic DNA is analyzed using DNA-analysis (Sanger et al. (1977), methods such as DNA sequencing. Proc. Natl. Acad. Sci. USA 74: 5463-5467)

Essential S.cerevisiae genes may thus be used for the identification of functionally similar genes and/or genes homologous in sequence in other mycetes, especially essential genes functionally similar and/or homologous in sequence in mycetes pathogenic to human, animal and plants. For this purpose for example mycetes of the classes phycomycetes or eumycetes may be used, in particular the ascomycetes, basidiomycetes, subclasses mehiascomycetales (yeast) and plectascales (mould fungus) and gymnascales (skin and hair fungus) or of the class of hyphomycetes, in particular the subclasses conidiosporales (skin fungus) and thallosporales (budding or gemmiparous among which particularly the species mucor, fungus), (blastomyces coccidioides, paracoccidioides rhizopus, aspergillus, (blastomyces), endomyces brasiliensis), penicilium (scopulariopsis), trichophyton (ctenomyces), hormodendron, piedraia, microsporon, epidermophton, cryptococcus, candida, sporotrichon, phialophora, geotrichum and trichosporon.

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Of particular interest is the use of Candida Spp. especially Candida albicans, Candida glabrata, Aspergillus Spp., especially Aspergillus fumigatus, Coccidioides immitis, Cryptococcus neoformans, Histoplasma capsulatum, Blastomyces dermatitidis, Paracoccidioides brasiliens and Sporothrix schenckii.

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Starting from the genes of S.cerevisiae, identified according to the above-described method, Applicants cloned corresponding essential genes from C.albicans i.e. CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, by the following method.

First, oligonucleotide(s) is(are) selected in the sequence of the S.cerevisiae gene or a homologous C. albicans sequence in order to amplify the corresponding fragment of C.albicans. After cloning, the obtained fragment (exhibiting a sequence of about several hundred bp) is used as a probe for screening a C.albicans (genomic) DNA library. The screening may include the following steps: clones were spread on dishes, covered with filters where the DNA was crosslinked to the filters, filters are hybridized, the positive colonies are then detected. The selected clone(s) is (are) then sequenced.

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The method contemplates that essential mycete genes are used to identify substances which may inhibit partially or totally the functional expression of these essential genes and/or the functional activity of the encoded proteins. Substances may be identified in this fashion, which inhibit mycetes growth and which can be used as antimycotics, for example in the preparation of drugs.

The present invention especially covers a method for screening such inhibiting substances wherein an essential gene from C.albicans selected from CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, or a

functionally similar gene in another pathogenic mycete or the corresponding encoded protein is used as target.

By functionally similar genes in other pathogenic mycete species, is meant genes which have a function similar or identical to that of the identified essential genes of C.albicans. Functionally similar genes in other pathogenic mycetes may, but need not be homologous in sequence to the corresponding essential C.albicans genes. Functionally similar genes in other pathogenic mycetes may exhibit only moderate sequence homology at the nucleotide level to the corresponding essential C.albicans genes. By moderate sequence homology it is meant in the present invention genes having a sequence identity, at the nucleotide level, of at least 50%, more preferably of at least 60% and most preferably of at least 70%.

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In addition, functionally similar genes in other pathogenic mycetes may, but need not encode proteins homologous in sequence to the proteins encoded by the essential C.albicans genes. Functionally similar proteins in other mycetes may exhibit moderate protein sequence homology to the proteins encoded by the essential C.albicans genes.

By moderate protein sequence homology is meant in the present invention proteins having a sequence identity, at the amino-acid level, of a least 40%, preferably of at least 50%, more preferably of at least 60% and most preferably of at least 70%.

A particular feature of this method is that essential mycete genes or the corresponding encoded proteins are used as targets for the screening of the substances. The method contemplates that essential C.albicans genes as well as functionally similar genes and/or genes homologous in sequence of other pathogenic mycetes or the corresponding encoded proteins may be used as targets.

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According to one embodiment of the screening method of the invention, mycetes cells are provided, which contain the essential gene used as target, and those cells are incubated with the substance to be tested. By this way, the growth inhibitory effect of this substance with respect to the essential target gene is determined.

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The mycetes cells which express the essential target gene to a different degree are used, and these cells are then incubated with the substance to be tested and the growth inhibitory effect of this substance is determined.

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The method includes the use of two or more mycetes cells, or strains derived therefrom, which differ in that they express the essential target gene to a different degree.

15 For example, two, three, four, five, ten or more mycetes cells or the corresponding mycetes strains may be comparatively analysed with respect to the growth inhibitory effect of a substance used in a defined concentration. Through such expression degree series, 20 antimycotic substances may be distinguished from cytotoxic or inactive substances.

A particular embodiment of the method includes the use of haploid mycetes cells/ strains for the screening, especially haploid S.cerevisiae cells/ strains.

25 The method contemplates the integration of the essential gene selected as a target in a suitable expression vector.

As expression vectors E.coli/S.cerevisiae shuttle vectors are for example suitable. Especially vectors differing in their copy number per cell may be used. Therefore, one may use vectors, which are present in the transformed S.cerevisiae cells in a high copy number, or one can also use those with a low copy number. One embodiment comprises the use of expression vectors which

allow the integration of the target gene in the S.cerevisiae genome.

For example the vectors pRS423, pRS424,pRS425, pRS426, pRS313, pRS314, pRS315, pRS316, pRS303, pRS304, pRS305, pRS306 (Sikorki and Hieter, 1989; Christianson et al. 1992) are appropriate as expression vectors.

The vectors of the series pRS423 - pRS426 are present in a high copy number, about 50 - 100 copies/ cell. On the contrary, the vectors of the series pRS313 - pRS316 are present in a low copy number (1 - 2 copies / cell). When expression vectors from these two series are used, then the target gene is present as an extrachromosomal copy. Using the vector of the series pRS303 - pRS306 allows the integration of the target genes into the genome. Using these three different expression vector types allows a gradual expression of the studied functionally similar essential gene.

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The method includes that the growth inhibitory effect of substances with respect to mycetes cells/strains is comparatively determined using expression vectors differing for instance in the copy number of the vector/ cell.

Such cells may express the essential target gene to a different degree and may exhibit a graduated reaction with respect to the substance.

target includes also, that The method expression of different strength is obtained in different mycetes cells (regulated overexpression) by insertion of the target gene in the expression vector between specific terminators. and S.cerevisiae promoters selected S.cerevisiae promoters which are constitutively expressed, but with different strength, are suitable. Examples for such promoters are native promoters of S.cerevisiae genes MET25, PGK1, TPI1, TDH3, ADH1, URA3, TRP1, as well as corresponding derivatives therefrom, for example promoter

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derivatives without specific activator and/or repressor sequences.

Regulated promoters are also appropriate for graduated over-expression of the target gene. The native promoters of the GAL1 genes and/or corresponding derivates thereof, for example promoters, in which different UAS elements have been eliminated. (GALS, GALL; Mumberg et al. (1994) Nucleic Acids Research 22: 5767-5768) as well as promoters of gluconeogenic genes, for example the promoters FBP1, PCK1, ICL1, or parts thereof, for example their activator- (UAS1 or UAS2) or repressor- (URS) sequences are used in corresponding non activable and/or non repressible test promoters (Schüller et al. (1992) EMBO J. 11: 107-114) Guarente et al. (1984) Cell 36: 503-511; Niederacher et al. (1992) Curr. Genet. 22: 363-370; Proft et al. (1995) Mol. 15 Gen. Genet. 246: 367-373).

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In the expression vector terminator for example the terminator sequence of S.cerevisiae genes MET25, PGK1, TPI1, TDH3, ADHI, URA3 may be used.

The method includes that by the use of cleverly 20 selected expression vector types and/or the preparation of suitable expression vectors, eventually using promoters of different strength and differently regulated promoters, a series of expression vectors may be constructed, containing the same target gene, but differing in that they express the target gene to a different extent.

the transformation of the method includes wild-type haploid vector in expression S.cerevisiae. The thus obtained S.cerevisiae cells/strains are cultivated in liquid medium and incubated in presence of different concentrations of the substance and the effect of this substance on the growth behaviour of the cells/strains expressing the target gene to a different degree is comparatively analysed. The method WO 00/15838 26

also includes that haploid S.cerevisiae cells/strains, transformed using the respective expression vector type without target gene, are used as a reference.

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The method includes that the screening of the substances can be carried out in different media using regulated promoters, especially GAL1 promoter and its derivates (GALS and GALL), since the expression degree may be largely influenced by the choice of the respective medium. Thus, the expression degree of the GAL1 promoter decreases in the following fashion: 2 % galactose > 1 % galactose + 1 % glucose > 2 % glycerine > 2 % glucose.

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The effect of the substances inhibiting the growth of wild-type cells of S.cerevisiae, may be partially or totally compensated by the overexpression of the functionally similar gene of another mycete species.

According to one embodiment, the method for screening antimycotic substances is carried out in vitro by contact an essential or functionally similar gene or the corresponding encoded protein with the substance to be tested and determination of the effect of the substance on 20 the target. Any in vitro test appropriate for determining the interaction of two molecules, such as a hybridization test or a functional test, can be used (e.g. enzymatic tests which are described in details in Bergmeyer H.U., Methods of Enzymatic Analysis, VCH Publishers). If the screening is carried out using the encoded protein as the target, then the corresponding essential gene is inserted by any suitable method known in the art, such as PCR amplification using a set of primers containing appropriate restriction sites, (Current Protocol in Molecular Biology, John Wiley and Sons, Inc) into an expression system, such as E. coli, Baculovirus, or yeast, and the expressed protein is then completely or partially purified by a Any purification method method known in the art.

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appropriate for the purification of expressed proteins, such as affinity chromatography can be used. If the target protein function is known, a functional test can then be carried out in which the effect of the antimycotic substance on the protein function is determined. If the protein function is unknown, substances which can interact with the target protein, e.g. which bind to the encoded protein, can be tested. In such a case a test such as protection of the target protein from enzymatic digestion by appropriate enzymes can be used.

According to one specific embodiment, the method for screening antimycotic substances corresponds to an enzymatic assay wherein the activity of dihydropteroate synthase (DHPS) and/or 7,8-dihydro-6-hydroxymethylpterin-pyrophosphokinase (HPPK) is determined; the enzymatic essay can be such as diclosed in "Bergmeyer H.U., Methods in Enzymatic analysis, VCH Publishers".

Dihydropteroate synthetase (DHPS) catalyses the 6-hydroxymethyl-7,8-dihydropterin of condensation pyrophosphate to para-aminobenzoic acid to form 7,8dihydropteroate which corresponds to the second step in the 6-hydroxymethyl-7,8from three-step pathway leading 7,8-dihydro-6-7,8-dihydrofolate. dihydropterin to hydroxymethylpterin-pyrophosphokinase (HPPK) catalyzes the 6-hydroxymethyl-7,8pyrophosphate to attachment of dihydropterin to form to 6-hydroxymethyl-7,8-dihydropterin pyrophosphate which corresponds to the first step in a three-step pathway leading to 7,8-dihydrofolate. All reduced folate cofactors for reguire organisms synthesis of a variety of metabolites. Most microorganisms must synthesize folate de novo because they lack the active transport system of higher vertebrate cells which allows these organisms to use dietary folates. Enzymes involved in folate biosynthesis are therefore targets for a variety

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of antimicrobial agents. Consequently, these enzyme activities are essential to the microorganisms, and are absent in man.

The method also includes the identification of genes which are functionally similar and/or homologous in sequence to essential C.albicans genes from humans, animals or plants. The corresponding human, animal or plant genes may optionally be used as target genes in the method in order to test if antimycotic substances exhibit an effect on these target genes.

A particular advantage of the method is that in this way substances may be identified which efficiently inhibit mycetes growth and also the influence of these substances on corresponding functionally similar genes and/or genes homologous in sequence to essential C.albicans genes from human, animal or plants may be determined.

The method includes also the possibility to check the existence of functionally similar genes and/or human, animal or plant genes homologous in sequence to the corresponding essential mycete genes, for example by checking homology of the identified essential mycete genes or parts thereof with human, animal or plant sequence genes available in data banks. In this way, it is possible to select at an early stage from the identified essential mycete genes, depending on the aim, those for which no functionally similar gene and/or no human gene homologous in sequence exist, for example.

Thereby, the method offers a plurality of possibilities to identify selectively substances with antimycotic effects, with no harmful effect on human beings.

For example, it is possible to identify substances usable for the preparation of drugs for the treatment of mycosis or prophylaxis in immunodepression states. These

substances may be used for example for the manufacture of drugs usable for the treatment of mycotic infections, which occur during diseases like AIDS or Diabetes. Substances which may be used for the fabrication of fungicides, especially of fungicides which are harmless for humans and animals, can also be identified.

Furthermore, the method offers the possibility to identify antimycotic substances, which selectively inhibit growth of specific mycete species only.

10 The screening method is particularly advantageous inasmuch as it is sufficient to know whether the genes are essential, one does not need any additional information regarding the function of the essential genes or the function of the encoded proteins. In addition, it is particularly advantageous for the identification of functionally similar genes to essential S.cerevisiae gene, in other mycetes where the DNA sequence is not available for many of these genes.

According to another aspect the invention provides an antibody directed against the protein encoded by the CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039, gene or a polypeptidic fragment thereof. The term "antibody" encompasses monoclonal and polyclonal antibodies. Said antibodies can be prepared by method well known in the art such as those disclosed in "Antibodies, a laboratory manual", Ed. Harbow and David Lane. Cold Spring Harbor Laboratory Eds., 1988.

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According to another aspect the present invention provides a kit for the diagnosis of fungal infections comprising a gene selected from the group consisting of CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 and CaJL039, a functionally similar gene thereof, a functional fragment thereof, the corresponding encoded protein, a functional polypeptide fragment thereof or an antiboby

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directed against the protein encoded by CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 or CaJL039 gene or by a functionally similar gene, or a polypeptidic fragment Such kits can be prepared using any suitable method well known in the art.

#### Examples

# Example 1 : CaNL256

Standford of site Internet (http://candida.standford.edu/) gives access to preliminary sequences of the genome of C. albicans. One of these 10 sequences has homology with the YNL256 gene of cerevisiae. Two oligonucleotides were selected in this (5'-ATTCATCCCATCAGTGCAGAAAG-3' and sequence in order to amplify ATTGACCAATAGCTCTAATTAATG-3') After cloning, we corresponding fragment of C. albicans. obtained a sequence of 399 bp close to the expected sequence (SEQ ID NO:1). The deduced protein was compared with the one of YNL256, evidencing 53% similarity and 43% identity (fig.1). This fragment of 399 bp of C. albicans was used as a probe for screening a genomic library of C. 20 albicans. The latter was prepared by partial digestion of genomic DNA of C. albicans by Sau3AI and cloning into the YEP24 vector at the BamHI site. The clones of the library were then spread at a density of 2000 clones per dish. Each dish was covered by a nitrocellulose filter which was then successively treated with: NaOH, 0.5M, 5 minutes; Tris, 1M, pH 7.7, 5 minutes; Tris, 0.5M, pH 7.7, Nacl, 1.25M, 5 minutes. After drying, the filters were kept for 2 hours at 80°C. Prehybridization and hybridization were carried out in a buffer of 40% formamide, 5xSSC, 20 mM Tris 30 pH 7.7 1xDenhardt 0.1% SDS. The probe was labeled with 32P with the RediPrime kit and dCTP from Amersham Hybridization took place over 17 hours at 42°C. filters were then washed in 1x SSC, 0.1% SDS, three times

for 5 minutes at room temperature and then twice for 30 minutes at 60°C, and were then submitted to autoradiography The colonies corresponding to the spots overnight. obtained were reisolated by re-spreading at low density followed by further hybridization. Three clones were thus obtained (out of 40,000), which were sequenced on an ABI 377 apparatus. The sequences were compiled using the ABI software and then analysed using the GCG software package. One of these three clones turned out to contain the complete coding sequence corresponding to the probe used; 10 this gene was called CaNL256, whose sequence is represented in SEQ ID NO:2. CaNL256 has 52% of nucleotides identical The coding region is shorter to YNL256 of S. cerevisiae. at the N-terminus. For translation to amino acids, account was taken of the fact that, in C. albicans, the CTG codon 15 is translated to Serine (there are 3 CTG codons The deduced protein had 40% amino acids CaNL256). identical with YNL256 of S. cerevisiae and 41% with FAS Pneumocystis synthase) of acid (folic Investigation into the databases using the Blast software 20 showed homology of two parts of the CaNL256 protein with, enzymes Dihydropteroate bacterial respectively, the Synthase (EC 2.5.1.15) (DHPS) of Haemophilus influenzae, meningitidis, haemolyticus, Neisseria Staphylococcus 25 Streptococcus pneumoniae, Bacillus subtilis, Clostridium acetobutylicum, Escherichia coli, Mycobacterium leprae (P 7,8-dihydro-6e-28) and than less value hydroxymethylpterin-pyrophosphokinase (EC 2.7.6.3) (HPPK) Haemophilus Escherichia coli, subtilis, Bacillus influenzae, Streptococcus pneumoniae (P value less than e-30 20). The units characteristic of DHPS and HPPK are also found in CaNL256.

# Example 2 : CaBR102

Stanford of site Internet The (http://candida.stanford.edu/) give access to preliminary sequences of the genome of C. albicans. One of these the YBR102 homology with has sequences S.cerevisiae. Two oligonucleotides were selected in this (5'-AGTATTCAATTGGGTATTCC-3' sequence amplified the to CCGGCATCATCAGTAACTCC-3') order in corresponding fragment of C. albicans. After cloning, we obtained a sequence of 647 bp (SEQ ID NO:3). The deduced 10 protein was compared with the one of YNL102, evidencing 35% similarity and 26% identity (fig.2). This fragment was amplified using Pfu polymerase (Stratagene). product was purified (High Pure PCR Product Purification Kit, Boehringer Mannheim) and used as a probe for screening 15 a C. albicans genomic DNA library. The latter was prepared by partial digestion of C. albicans genomic DNA with SauIIIA and cloning into the YEP-24TRP1 vector at the BamHI restriction site. 40,000 clones of the library were then spread at a density of 2000 clones per dish. Each dish was 20 covered by a nitrocellulose filter (Membrane Hybond  $N^*$ , Amersham) which was then successively treated with: 1.5 M NaCl/0.5 M NaOH, 5 minutes; 1.5 M NaCl/0.5 M Tris-HCl pH 7.2/1 mM EDTA, 3 minutes, twice; DNA was crosslinked to the filters (Amersham Life Science, ultra violet crosslinker). 25 The probe (100 ng) was labelled with 32P using the RediPrime kit and dCTP (Amersham Life Science). The filters were hybridized in a buffer containing 30% formamide, SSC, 5% Denhart's solution, 1% SDS, 100  $\mu g$  /ml salmon sperm DNA and a probe concentration of 106 cpm/ml at 42°C for 16 The membranes were then washed three times at room temperature in 2 x SSC/0.1% SDS for 5 minutes each and three times in 1 x SSC/0.1% SDS at 60°C for 20 minutes each. the filters were then exposed overnight to an X-ray

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film. The colonies corresponding to the positives clones were isolated and screened a second time by the same procedure. Two positives clones were finally obtained, which were sequenced on an ABI377 apparatus. the sequences were compiled using ABI software and then analysed using the GCG software package. The nucleotide sequences of these two clones were identical and contained the complete coding sequence corresponding to the probe used, this gene was called CaBR102, whose sequence is represented in SEQ ID The translation of this nucleotide sequence was No:4. examined, account was taken of the fact that in C. albicans the CTG codon is translated to serine (there are 3 CTG codons in CaBR102). The deduced protein has 24% identity to S. cerevisiae gene YBR102.

# Example 3 : CaIR012

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Chromosomal DNA from the C. albicans strain Caf2-1 was isolated using Yeast Cell Lysis prep Kit and Genome DNA Kit A 343 bp fragment from C. albicans genomic from BIO101. DNA (SEQ ID NO:5) was amplified with the oligonucleotide primers CaYIR012-5' (5'-GACGTCGTAGACGATACTCAAGAAG-3') (5'-CTGCAGTAAACCCTCCAGATATAACAG-3') CaYIR012-3' PowerScript DNA polymerase (PAN Systems GmbH) using the hot start technique. The PCR product was purified agarose gel and labeled with fluorescein (Gene image random prime labelling module, Amersham Life Science) according to the manufacturer's instructions. Plasmid DNA from E.coli was isolated using Qiagen columns as recommended by the Screening the  $\lambda ZAPII$  C. albicans cDNA manufacturer. manufacturer's the performed following library was Nylon Ltd.). (Stratagene instructions (Schleicher&Schuell) were lifted from LB-plates (150 mm) with 15000 pfu/plate, denatured 5 min in 1.5M NaCL, 0.5M NaOH, neutralized 3 min in 1.5M NaCl, 0.5M Tris-HCl pH8.0, washed 3 min in 0.2 M Tris-HCl pH 7.5, 2xSSC and DNA was

crosslinked to the filters (Stratagene UV crosslinker). The filters were prehybridized 4 h at 60 °C and hybridized with the fluorescein-labeled DNA probe overnight at 60 °C. Detection was performed with Anti-fluorescein AP conjugate (Signal amplification module for the FluorImager, Amersham LIFE SCIENCE) and analysed after 20 h with a Fluorimager (Storm 860, Molecular Dynamics). Positive plaques were picked and incubated with 0.5 ml SM-buffer (100mM NaCl, 8 50 mM Tris-HCl pH7.5, 0,01% gelatin). mM MgSO., selected clones were diluted, titered with host cells XL1-10 Blue and screened and purified a second time by the same phagemid SK(-) pBluescript procedure. Finally, the containing the DNA insert of interest was rescued by the ExAssist Helper Phage system according to the Stratagene protocol. From a total of 75000 screened plaques, 3 15 positive clones were identified. pBluescript SK (-) phagemid DNA was isolated, sequenced with T3 and T7 primers and the sequences were extended with custom-synthesized oligonucleotide primers. Nucleotide sequence analyses were performed with the Gene Data software package (Gene Data 20 Basel Switzerland). Similarity searches with the Swissprot database were conducted with the BLAST program (Gish, Warren and David J. States (1993). Identification of protein coding regions by database similarity search. Nat. Genet. 3:266-72.). One of these three clones turned out to contain the complete coding sequence corresponding to the probe used; this gene was called CaIR1012, whose sequence is represented in SEQ ID NO:6.

# Example 4: CaJL039

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The CaJL039 sequence is depicted in SEQ ID No 7.

The CaJL039 gene was cloned based on gene fragment data issued from the public Stanford Candida albicans sequencing database.

(a) A fragment that showed homology to Saccharomyces cerevisiae YJL039c was identified, the sequence of which is given in SEQ ID No 8.

Using the procedure disclosed in example 3 with the oligonucleotide primer pair (Ca039s: TAG CTC AAC CTA CCA CCA ATC /Ca039r: ATC ACA AGA CTG TCA ATG TAA AT), a short PCR fragment (234 base pairs long) was amplified for screening a Candida albicans cDNA lambda ZAP II library (gift of Alistair Brown, Aberdeen).

Three positive clones of the 3' coding region were 10 obtained.(# 21t7, 11t3, 21t3).

3'- and 5'- extension of the internal (b) fragment using the primer walking method

The Sanglard genomic Candida DNA library with the YEp24 vector backbone was used for further amplification of 15 3'- and 5'-coding sequences. Amplification was carried out by using the following vector-specific oligonucleotide primers and CaJL039 fragment-specific primers:

cggaattcctatcgactacgcgatcatgg: YEp24for (vector

specific) 20

> gcgaattccgatataggcgccagcaac: YEp24ba (vector specific)

caattgctttgactcgggtgttattaagt: Ca039-51 (CaJL039: 5'fishing)

tcttggcacaacttgataagaatctgt: Ca039-52 (`) 25

taggtgtacgcgaaagccaagtagaac:Ca039-53 (`) ttgttaatcgtacacctaaggtgttgac: Ca039-31 (CaJL039: 3'fishing)

ttgcagattgatgctagcaatgtatttg: Ca039-32 (`)

Using the technique of primer walking, the complete 30 5'-sequence could be amplified (clone 14b-1-1 and clone 17b-3-4).

3'-sequence was available from GTC The missing PathoGenome Release 5.0, contig #2830.

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An interacting protein (C82, component for RNA polymerase III in yeast) has been identified.

#### Example 5: CaOR110

#### 5.1. CaOR110

The CaOR110 sequence is depicted in SEQ ID No 9.

CaOR110 was cloned based on gene fragment data issued from the public Stanford Candida albicans sequencing database.

- (a) A small ScOR110-homologous fragment was used in a hybridization experiment to identify CaOR110 clones in a Candida Albicans lambda ZAPII cDNA library (from Alistair Brown). Alignement of Candida Albicans CaOR110 sequence with the fragment used for hybridization is given in figure 3. The homologous fragment sequence is given in SEQ ID No. 15
  - (b) 3'- and 5'- Extension of the internal fragment:

The Sanglard genomic Candida DNA library (received from RMV) in the YEP24 vector backbone was used for the amplification of 3'- and 5'- coding and non-coding sequences. This amplification was done by using the vector-specific oligos (directional towards the insert) and CaOR110 fragment-specific oligos (directional towards the vector flanking sequences) described below:

cggaattcctatcgactacgcgatcatgg : YEP24for

- 25 gcgaattccgatataggcgccagcaac : YEP24ba
  - cgggatccggtaaccaattggatctataaccgtg: 110-ba-150
    gcggatcctggtgcccttggtggtgaatg: CaYOR110A
    gcggatccctcacaatatgacgattgaaact: CaYOR110B
    ggcgtcgactcaggcgccagttttacgtacttcaaattcatc: CaYOR110C
- tgtgaattcttgacacagggtga : CaYOR110D caaaccttcagcacaactcca : CaYOR110E

The finaly assembled sequence that included also 3'- and 5'- non-coding sequences was verified by sequencing. The coding region was subcloned into the p414RSGALL-vector.

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The map is depicted in Fig. 4.

The homologous yeast ORF (YOR110w) has been described as the transcription factor subunit TFC7 interacting with TFC1 in the TFIIIC polymerase complex (Manaud et al., 1998, Mol. Cell. Biol. 18; 3191-3200).

#### 5.2. CaOR110 splice variant

For CaOR110, an additional splice variant was identified. The clones for the splice variant of CaOR110 were obtained from a Candida albicans cDNA library.

The sequence is depicted in SEQ ID No.10.

The splice variant uses the donor site "gtacgt" at position 907 of the original CaOR110 sequence. Acceptor site is at 1047. The map is disclosed in Fig. 5.

The alignement of the original CaOR110 and the splice variant is given in fig. 6.

#### Example 6 : CaMR212

The CaMR212 sequence is depicted in SEQ ID No. 11.

(a) CaMR212 was cloned based on gene fragment data from the public Standford Candida albicans sequencing database.

The sequence of a fragment showing homology (Blast search) to the Saccharomyces cerevisiae gene YMR212c is given in SEQ ID 12.

Based on these data, the following oligos were designed that allow amplification of this fragment (490 bp-fragment) from genomic Candida albicans DNA.

Oligos:

CaYMR212for: 5'- cacctgtgaacaacccaccatc-3'
CaYMR212back: 5'- gaatatcctttttaactcaagag -3'

(b) 3'- and 5'- extension of this internal fragment 30 from CaMR212

For this purpose, genomic Candida DNA libraries from Dominique Sanglard (received from RMV) were used. The YEp24 backbone of this library was used to amplify the 3'- and 5'- coding and non coding sequences with PCR. This was done

by using oligos specific for the CaMR212 490 bp-fragment (directional towards the vector flanking regions) and vector-specific oligos (directional towards the insert).

Oligos:

5

YEP24for (vector specific):

5'-cggaattcctatcgactacgcgatcatgg

YEP24ba (vector specific):

5'-qcgaattccgatataggcgccagcaac

Primer YEp24for and CaMR212for gave a 500 bp fragment, 10 encoding 5'-UTR and the 5'coding region from CaMR212.

Using primer YEp24 for and CaMR212back a 1400 bp CaMR212-fragment was amplified. Using the sequence of this 1400 bp-fragment the following new primers, specific for this fragment were designed.

15 Oligos:

Ca212-1: 5'- gctttcccagcaggataacattg

Ca212-2: 5'- tgagttataatgcagctgttgg

Ca212-3: 5'- catctcgtgtgaacatgattgg

Primers YEP24 for and Ca212-3 gave a 1600 bp fragment, coding for the 3´- coding region and the 3´UTS region.

With the 3 PCR fragments the 2900 bp sequence(including coding and 3'and 5'-non-coding sequences) was assembled. With the following new primers the coding sequences was amplified from genomic DNA and cloned into

25 p413GALL-vector.

Oligos for amplifying coding region:

Ca212for: 5'- agtttcttcaacttccagatccaag

Ca212back: 5'- gtatatttgcaactgtctctctctc

The yeast homolog YMR212c plays a role in cell wall function because the knockout can be rescued in 1M sorbitol. In addition, YMR212c unger GAL-promoter regulation shows an increased sensitivity versus Congo Red and Calcofluor White. YMR212c is an integral membrane protein and localizes to the plasma membrane (demonstrated

by microscope analysis of YMR212-GFP fusion proteins and by biochemical analysis of YMR212-GST fusion proteins).

#### Example 7 : CaDR325

The CaDR325 sequence is given at SEQ ID 13.

CaDR325 was cloned based on gene fragment data from 5 the public Stanford Candida albicans sequencing database.

- (a) 3 fragments that showed homology to Saccharomyces cerevisiae YDR325 were identified, the sequences of which are disclosed in SEQ ID 14, 15 and 16.
- Based on these data, the following oligos were 10 designed that allowed the verification of the database sequences and the amplification of an approx. 2200 bp internal CaDR325 fragment from genomic DNA:

cgagcatctacttgttcaaccac: hybCaYDR325ba Oligo

gaatctctggctcgctc: 15

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325-juls Oligo

gaccgagatacacgagaat: 325-julr Oligo

ggttaaatgatcgtgatgaat: Ca325r Oligo

caacctcactgacaaatactt: Ca325s Oligo

The finally subcloned 2200 bp internal fragment was amplified by the combination hybCaYDR325ba + 325-julr oligos.

(c) 3'- and 5'- Extension of the internal fragment:

The Sanglard genomic Candida DNA library (received from RMV) in the YEP24 vector backbone was used for the amplification of 3'- and 5'- coding and non-coding sequences. This was done by using the following vectorspecific oligos (directional towards the insert) CaDR325 2200 bp fragment-specific oligos (directional towards the vector flanking sequences):

YEP24for (vector cggaattcctatcgactacgcgatcatgg : 30 specific)

(vector gcgaattccgatataggcgccagcaac : YEP24ba specific)

acgcttccaatgtattattctcg : Oligo 1-10-A back

40

ggatgccaatttccctga: Oligo 1-10-B for catccagaagatataacggct: Oligo 1-10-C for tgcataatctactcagcgaca: Oligo 1-10-D back gtggttgaacaagtagatgctcg: Oligo 1-10-E for caggttgaacaagtagatggattg: Ca325Klon\_2\_Fo

gcgcttgaaaccactagtgaattg : Ca325Klon\_2\_Fo caattcactagtggtttcaagcgc : Ca325Klon\_3\_Ba

The finally assembled 4700 bp sequence that included also 3'- and 5'- non-coding sequences were verified by sequencing. The coding region was subcloned into the p413RSGALL-vector.

The map is disclosed in fig. 7.

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Sequences numbers are identified in field 130 of the sequence listing.

Claims:

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1. A polynucleotide having the sequence as depicted in SEQ ID No.2, SEQ ID No.4, SEQ ID No.6, SEQ ID No.7, SEQ ID No.9, SEQ ID No.10, SEQ ID No.11 or SEQ ID No.13, homologs thereof and functional fragments thereof.

- 2.-The polynucleotide of claim 1 having the sequence as depicted in SEQ ID No.2, SEQ ID No.4, SEQ ID No.6, SEQ ID No.9, SEQ ID No.10 or SEQ ID No.11, homologs thereof and functional fragments thereof.
- 3.-The polynucleotide of claim 1 which is the gene CaNL256, homologs thereof and functional fragments thereof.
- 4.-The polynucleotide of claim 1 which is the gene CaBR102, homologs thereof and functional fragments thereof.
- 5.-The polynucleotide of claim 1 which is the gene 20 CaIR012, homologs thereof and functional fragments thereof.
  - 6.-The polynucleotide of claim 1 which is the gene CaMR212, homologs thereof and functional fragments thereof.
- 7.-The polynucleotide of claim 1 which is the gene CaDR325, homologs thereof and functional fragments thereof.
  - 8.-The polynucleotide of claim 1 which is the gene CaOR110, homologs thereof and functional fragments thereof.
  - 9.-The polynucleotide of claim 1 which is the gene CaJL039, homologs thereof and functional fragments thereof.

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10.-A gene according to any one of claims 3 to 9, wherein the functionally similar gene has a sequence identity, at the nucleotide level, of at least 50%, preferably of at least 60%, and most preferably of at least 70%.

- 11.-A gene according to any one of claims 3 to 9, wherein the functionally similar gene has a sequence identity, at the amino-acid level, with the encoded protein, of at least 40%, preferably of at least 50%, more preferably of at least 60% and most preferably of at least 70%.
- 12 -A protein encoded by the polynucleotide according 15 to any one of claims 1 to 11, a functional polypeptide fragment thereof.
  - 13.-A plasmid containing the gene according to any one of claims 3 to 9.

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- 14.-A plasmid deposited at the CNCM with the accession number I-2065.
- 15.-A plasmid deposited at the CNCM with the accession number I-2063.
  - 16.-A plasmid deposited at the DSMZ with the accession number DSM 12977.
- 17.-A plasmid deposited at the DSMZ with the accession number DSM 12976.
  - 18.-A plasmid deposited at the DSMZ with the accession number DSM 12978.

- 19.-A plasmid deposited at the DSMZ with the accession number DSM 12979.
- 5 20.-A polynucleotide obtainable by the process comprising the following steps:
  - (i) selecting an essential gene from Saccharomyces cerevisiae;
- (ii) comparing the sequence of said gene with
  10 Candida Albicans genome sequences;
  - (iii) deducing homologuous oligonucleotides
    regions;
  - (iv) PCR amplifying the thus-obtained
    oligonucleotides;
- 15 (v) using the amplimers of step (iv) for detecting the complete gene of interest;

and homologs thereof and functional fragments thereof.

- 21.-The polynucleotide of claim 20, in which step (v)
  20 is comprised of the step of using the amplimers of step
  (iv) as a probe for detecting the complete gene of interest
  from a Candida albicans genomic library.
- 22.-The polynucleotide of claim 20, in which step (v)
  25 is comprised of the step of using the amplimers of step
  (iv) as a probe for detecting the complete gene of interest
  from a Candida albicans cDNA library.
- 23.-The polynucleotide of claim 20, in which step (v) 30 is comprised of the step of 3' and 5' extension of the amplimer using a PCR method.
  - 24.- An antiboby directed against the protein of claim 12 or a functional polypeptide fragment thereof.

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- 25.-A method for the screening of antimycotic substances wherein an essential gene from mycetes or a functionally similar gene from another pathogenic mycete, or the corresponding encoded protein, is used as target and wherein the essential gene is according to any one of claims 3 to 9.
- 26.-The method of claim 25 wherein mycete cells which

  10 express the essential gene, or a functionally similar

  mycete gene, to a different level are incubated with the

  substance to be tested and the growth inhibiting effect of

  the substance is determined.
- or the corresponding target encoded protein is contacted in vitro with the substance to be tested and the effect of the substance on the target is determined.
- 28.-The method according to any one of claims 25-27 wherein the screened substances partially or totally inhibit the functional expression of the essential genes or the functional activity of the encoded proteins.
- 29.-The method according to any one of claims 25-28 wherein the screened substances partially or totally inhibit the activity of dihydropteroate synthase (DHPS) and/or 7,8-dihydro-6-hydroxymethylpterin-pyrophosphokinase (HPPK).

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30.-The method according to any one of claims 25-29 wherein the mycete species are selected from the group comprising Basidiomycetes, Ascomycetes and Hyphomycetes.

- 31.- The method according to any one of claims 25-30, wherein said functionally similar genes are essential genes from Candida Spp, or Aspergillus Spp.
- 5 32.- The method according to claim 31, wherein said functionally similar genes are essential genes from Candida albicans, or Aspergillus fumigatus.
- 33.- The method according to any one of claims 25-32 wherein the functionally similar gene has a sequence identity, at the nucleotide level, with the corresponding essential gene of at least 50%, preferably of at least 60%, and most preferably of at least 70%.
- wherein the functionally similar gene encodes a protein having a sequence identity, at the amino-acid level, with the corresponding essential gene encoded protein of at least 40%, preferably of at least 50%, more preferably of at least 70%.
  - 35.- A kit for diagnosis of fungal infections comprising a gene selected from the group consisting of CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 and CaJL039, a functionally similar gene thereof, a functional fragment thereof, the corresponding encoded protein or a functional polypeptidic fragment thereof, or an antiboby directed against the protein encoded by the gene selected from the group consisting of CaOR110, CaMR212, CaNL256, CaBR102, CaIR012, CaDR325 and CaJL039, or by a functionally similar gene, or a polypeptidic fragment thereof.

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## Fig.1

1	IHPISAESLHSHLQQLINDKPQ	22
	:[]:-[] : -[]-[]	
451	PDLNIPHPRMLERTFVLEPLCELISPVHLHPVTAEPIVDHLKQLYDKQHD	500
23	ETVQESSDLLQFIPVSRLPVKDNILKFDQINHKSPTLIMGIL	64
501	EDTLWKLVPLPYRSGVEPRFLKFKTATKLDEFTGETNRITVSPTYIMAIF	550
65	NMTPDSFSDGGKHFGKELDNIVKQA.EKLVSEGATIIDIGGVSTRPG	110
551	NATPDSFSDGGEHFADIESQLNDIIKLCKDALYLHESVIIDVGGCSTRPN	600
111	SVEPTEEEELERVIPLIRAIRQS	133
	:: .    :       :  :	
- 0 1	CTOACEFEETBRGIDLIKATRESTELPODKVILSIDTYRSNVAKEAIKVG	650

### Fig.2

	NDLNEVLDQCTKIAEKRLQLQDQIDQERQGNFNNVESHSNSPALLPPLKA	300
251	MDDMEV DDQC I KIREKRDQDQDQIDQDKQQMM MAY 20110M911—2011	
1	ksiql	5
301	GONGNLMRRDRSSVLILEKFWDTELDQLFKNVEGAQKFINSTKGRHILMN	350
6	GIPSN.KKKDRSSIMVLKKMWDSQLQSLFKHVDGASKFVQPLPNRHIVAE	54
351	SANWMELNTTTGKPLQMVQIFILNDLVLIADKSRDKQNDFIVSQCYP	397
55	SGRWFEVNVGNWKPSYPTHLFIFNDLILIAVKKSSSSQEPTTGGSNGGS	104
398	LKDVTVTQEEFSTKRLLFKFSNSNSSLYECRDADECSRLLDVIRKAKD	445
105	KSRLQAVQCWPLTQVSLQQIKSPKKDDDKMYFINLKSKSLSYVYLTDRYD	154
446	DLCDIFHVEEENSKRIRESFRYLQSTQQTPGRENNRSPNKNKRRSMGG	493
155	HFVKVTEAFNKGRNEMIQSERLLDSRLSSPSNNNGDSKEEKRQLRESLRN	204
494	SITPGRNVTGAMDQYLLQNLTLSMHSRPRSRDMSSTAQRLKFLDEGVEEI	543
205	-	225

1G. :	3
301	${\tt ACCCATTGCTGAAATGTTGGACTTGAAGATTGCTTTAGAAAGAGGAGTTGGTGAATGGTT}$
0	
361	${\tt TCGTAAAAATAGAGATACCAAACCAGTTCCCGGTGATTACACACAATTGAGAACATTTTT}$
0	
	${\tt CGATAAATTATTGATCGATGAAGATACTTGGCCAAGAGATAACTTAAATGTTATACCTAA}$
0	
481	${\tt TATTGAAGGAGAAGATTATGATGAAAATCTACGATCGTGCCAAATTGTTTTGGAAAAAGTT}$
0	TTAAATATGTGTTGATAGTTACACATGC
541	THE TAX AS A REPORTED A A ATTA A A A ATCTCTTCATACTTACACATACAT
29	AGCAACGAAAATTGCTTTAGGATCAGCTTTATTACAGTTAAAATCAGTTACTGATGTTAT
601	AGCAACGAAAATTGCTTTAGGATCAGCTTTATTACAGTTAAAATCAGTTACTGATGTTAT
89	AGATGATAATCAAACTGTGTTACGTGCTGGTGCATGTTCATTATCCAAATTTGTTAGAGA
661	AGATGATAATCHAACTGTGTTACGTGCTGGTGCATGTTCATTATCCAAATTTGTTAGAGA AGATGATAATCAAACTGTGTTACGTGCTGGTGCATGTTCATTATCCAAATTTGTTAGAGA
149	TGGCGAAGATAAAACCAATCATACTATTCAATGGAAAATTGTCATGAATGGTAATTGTGA
721	TGGCGAAGATAAAACCATCATGAATGGAAAATTGTCATGAATGGTAATTGTGA
	ATTCTTGACACAGGGTGAAGAATGAAT
781	ATTCTTGACACAGGGTGAAGAAATGAACTGGGATTTCCGTCGTGGTGTTGAAGCCGGGTC

FIG.4

HinD III 287 Msc I 952 Drd I 290 Hae I 952 Xho I 295 Pvu I 1015 PacR7 1 295 BsiE I 1015 Ava I 295 AlwN 11151 Spe I 506 BspH I 1263 Hpa I 7 Xho II 617 EcoR 1 1280 Bcl I 57 BstY 1617 Pvu II 1343 Ban I 76 BstE II 627 NspB II 1343 Sty 181 Bsa I 793 MspA111343 Sec | 81 Mme I 799 Bgl 1 1448 BsaJ I 81 Xmn I 858 SnaB 1 1487 Ear I 103 BseR I 858 Psp1406 I 1544 BsrG I 233 Eae I 950 Afl III 1610

CaOR110

FIG.5

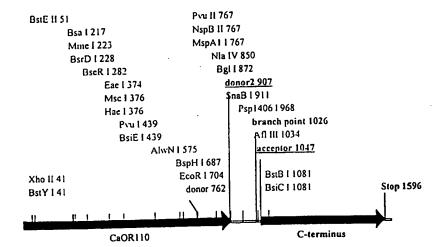


FIG.6

	ATGACGATTGAAACTATTTATATCGCAAGACACGGTTATAGATCCAATTGGTTACCACCA	60
1	ATGACGATTGAAACTATTTATATCGCAAGACACGGTTATAGATCCAATTGGTTACCACCA	60
	CCACACCCACAAATCCTACTGGTATTGACAGTGACCCGGCTTTAGCACCACATGGTGTT	120
61	CCACACCCACCAAATCCTACTGGTATTGACAGTGACCCGGCTTTAGCACCACATGGTGTT	120
	GAACAAGCCCAACAGTTAGCTGCCTATCTTACATCATTACCTACACATGAAAAGCCTGAA	180
121	GAACAAGCCCAACAGTTAGCTGCCTATCTTACATCATTACCTACACATGAAAAGCCTGAA	180
	TTTATTATTGCTTCACCTTTTTATCGTTGTATAGAAACGTCGAGACCCATTGCCGAAATG	240
181	TTTATTATTGCTTCACCTTTTTATCGTTGTATAGAAACGTCGAGACCCATTGCCGAAATG	240
	TTGGACTTGAAGATTGCTTTAGAAAGAGGAGTTGGTGAATGGTTTCGTAAAAATAGAGAT	300
	TTGGACTTGAAGATTGCTTTAGAAAGAGGAGTTGGTGAATGGTTTCGTAAAAATAGAGAT	300
	ACCAAACCAGTTCCCGGTGATTACACACAATTGAGAACATTTTCGATAAATTATTGATC	360 360
	ACCAAACCAGTTCCCGGTGATTACACACAATTGAGAACATTTTTCGATAAATTATTGATC	300
	GATGAAGATACTTGGCCAAGAGATAACTTAAATGTTATACCTAATATTGAAGGAGAAGAT	420 420
	GATGAAGATACTTGGCCAAGAGATAACTTAAATGTTATACCTAATATTGAAGGAGAAGAT	120
	TATGATGAAATCTACGATCGTGCCAAATTGTTTTGGAAAAAGTTTATTCCTGAATTTGAA	480
	TATGATGAAATCTACGATCGTGCCAAATTGTTTTGGAAAAAGTTTATTCCTGAATTTGAA	
,	AAGAAATTCCCCGAAATTAAAAATGTGTTGATAGTTACACATGCAGCAACGAAAATTGCT	540 540
	AAGAAATTCCCCGAAATTAAAAATGTGTTGATAGTTACACATGCAGCAACGAAAATTGCT	
	TTAGGATCAGCTTTATTACAGTTAAAATCAGTTACTGATGTTATAGATGATAATCAAACT	600
	GTGTTACGTGCTGGTGCATGTTCATTATCCAAATTTGTTAGAGATGGCGAAGATAAAACC	660
	GTGTTACGTGCTGGTGCATGTTCATTATCCAAATTTGTTAGAGATGGCGAAGATAAAACC	
661	AATCATACTATTCAATGGAAAATTGTCATGAATGGTAATTGTGAATTCTTGACACAGGGT	720

661	7 / 9 AATCATACTATTCAATGGAAAATTGTCATGAATGGTAATTGTGAATTCTTGACACAGGGT	720
		780
	GAAGAAATGAACTGGGATTTCCGTCGTGGTGTTGAAGCCGGGTCAGCTGAAGATATAGCG	780
781		840
781	CAAAGAAAGCAGCAGCAGAAGCAGAAGCATGAAGAAAATGAACAAACCAAA	840
841	TCCGATGGTCCCATCACTGAATCTGCCACTGGGGCAGAAATAGATGGGAATGAAGATGAA	900
841	TCCGATGGTCCCATCACTGAATCTGCCACTGGGGCAGAAATAGATGGGAATGAAGATGAA	900
901	TTTGAAGTACGTAAAACTTGAAAGAGATATTAAATAGACACAAACTTAGAAAATATAGAG	960
901	TTTGAA	906
961	$. \\$ ATACAAACGTTTTGAATTCTTGATTCACTTTTTTGTTTAAAAATAAAATAGTTCAAAA	1020
906		905
1021	TGAAATACTAACACATGTGTTTTTTAGACATTTTATGTAACCATCGATATACCTTCAATTT	1080
906		939
	CGAATAAAATCGACAATGAAGAAGAACCACCATCAAGGACAGGTCAAGCTCCAAAATTCA	1140
941	CGAATAAAATCGACAATGAAGAAGAACCACCATCAAGGACAGGTCAAGCTCCAAAATTCA	1000
	AAAACAATATTATCAAGCCTTCAGCACAACTCCAATTTACTGATTTAAAAGAAGATCATC	1200
1001	AAAACAATATTATCAAGCCTTCAGCACAACTCCAATTTACTGATTTAAAAGAAGATCATC	1060
1201	CATTAGTAAAAATATCGAACAATACTATATCTGCTCAAGGCTCGTCGTCGTCGTCGTTAT	
1061	CATTAGTAAAAATATCGAACAATACTATATCTGCTCAAGGCTCGTCGTCGTCGTTAT	1120
	CAGCGTCGAAAAATGGATTTAATAGTCATACTCACAATTCAGGAGTCATTGATCCATCAG	
1121	CAGCGTCGAAAAATGGATTTAATAGTCATACTCACAATTCAGGAGTCATTGATCCATCAG	1180
	CACTTATAGATGGGAAAATTTATCAGACTGATTGGAATCAATTACAAGGTACTGAACTAA	
1181	CACTTATAGATGGGAAAATTTATCAGACTGATTGGAATCAATTACAAGGTACTGAACTAA	1240
	TATTTGATGAAAATGGTCAATTTATAGGCAAGGTTAAGGAACATTTGACTTGCAATAATA	
1241	TATTTGATGAAAATGGTCAATTTATAGGCAAGGTTAAGGAACATTTGACTTGCAATAATA	1300

		•	•	•			
1421	AAAGAGCAA'	TTGTGGCTGC	TAGAGCCAAA	\GGTAA			1454
		[	1111111111	11111			
1561	AAAGAGCAA'	TTGTGGCTGC	TAGAGCCAAA	GGTAAATAAA	TGCTATTITG	ATATTATTATT	1020
		•	•			•	1620
	-	=	-				
1301	CIMICALGG	0	0	0	•	0 0	
1261	CTATCATCC	ነነነነነነነነነ ልጥልጥልርልጥርል	AGACTCACAA	GGACAACAAC	CAGCTAGAAG	TCAGTTCTTAA	1420
	1111111111	11111111111		111111111	11111111111	111111111	
1501	CTATCATGG	ATATAGATCA	AGACTCACAA	GGACAACAAC	CAGCTAGAAG	TCAGTTCTTAA	1560
			•		•	•	
	,	0	•	•	•		
1301	ACACAAAAT	TCACATTAAA	<b>LAAAGGCAGAA</b>	GAAGTAGAAC	AACTTCGTTC	AGCAGATGATT	1360
	111111111	!	1111111111	11111111111	111111111	11111111111	
1441	ACACAAAAT'	TCACATTAAA	AAAGGCAGAA	GAAGTAGAAC	AACTICGITC	AGCAGATGATT	1500
		•	•				7 5 0 0

PCT/EP99/07376

FIG.7

BsaJ 1 2587 BsaA 1 1203 Xho I 2623 Hac 1 1668 Bsm I 1758 PaeR7 1 2623 Ava 1 2623 , EcoR V 2207 Hpa I 2929 Hac II 2304 Hind II 2929 Pvu 11-78 Spc 1 2313 HinC II 2929 NspB 11 78 Msl 1 2362 HgiA 1 2558 Mme I 3308 MspA1 I 78 Pst 1 3476 Bsp1286 1 2558 BspH I 396 HinD III 3748 BsiHKA I 2558 AN 11 597 Xho II 4013 Sty I 2587 AN III 650 BstY 1 4013 Sec I 2587 BspM I 807 Bgl 11 4013 Nco I 2587 Psp 1406 1 887 Cla I 4022 Dsa 1 2587 SnaB I 1203

CaDR325

#### 1183

#### SEQUENCE LISTING

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gaactagata atattgtgaa gcaggcagag aaattagtca gtgagggtgc tacgattatt 300
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Tyr Lys Asn Val

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						gcc Ala											1488
						tct Ser											1536
						ttc Phe											1584
						act Thr											1632
(						caa Gln 550											1680
						cgt Arg											1728
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gta Val	tca Ser	aaa Lys	tta Leu 660	att Ile	gtt Val	ggt Gly	tta Leu	caa Gln 665	gga Gly	tca Ser	acg Thr	aaa Lys	ile 670	gat Asp	gtt Val	2016
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Asn	Leu	Leu	Gln	Val 85	Pro	Gly	Ala	Gly	Gly 90	Gly	Gly	Asp	Leu	Asn 95	Ser
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		115					120					125	Ala		
	130					135					140		Val		
Ala 145	Gly	Gly	Asp	Val	Met 150	Thr	Arg	Thr	Gly	Gly 155	Leu	Thr	Ile	Glu	Gln 160
•				165					170				Val	175	
Tyr	Tyr	Lys	Thr 180	Leu	Leu	Lys	Glņ	Lys 185	Asn	Leu	Ile	Thr	Arg 190	Asp	Ile
· **		195					200					205	Leu		
	210					215					220		Gly		
225					230					235			Ala		240
				245	٠				250				Leu	255	
Pro	Gln	Lys	Ser 260		Gln	Leu	Gly	Ile 265	Pro	Ser	Asn	Lys	Lys 270	Lys	Asp

Arg	Ser	Ser 275	Ile	Met	Val	Leu	Lys 280	Lys	Met	Trp	Asp	Ser 285	Gln	Leu	Gln
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Leu	Ile	Ser	Ser 740	Gly	Ile	Asn	Met	Glu 745	Thr	Asp	Glu	Pro	Ile 750	туr	Lys
Val	Lys	Glu 755		Lys	Leu	Tyr	Asp 760		Phe	Leu	Lys	Ile 7 <b>6</b> 5	Met	Gln	Pro
Gln	Leu 770		Glu	Leu	Lys	Leu 775		Gly	Leu	Asn	Val 780		Tyr	Ile	Phe

Glu Ser Ile Leu Asn Leu Glu 785 790

PCT/EP99/07376

### SEQUENCE LISTING

- <110> Hoechst Marion Roussel
- <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES
- <130> SEQID05
- <140>
- <141>
- <160> 1
- <170> PatentIn Ver. 2.1
- <210> 1
- <211> 343
- <212> DNA
- <213> Artificial Sequence
- <223> Description of Artificial Sequence:probe

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### 22/23

### SEQUENCE LISTING

<110> Hoechst Marion Roussel <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES <130> SEQID06 <140> <141> <160> 2 <170> PatentIn Ver. 2.1 <210> 1 <211> 1248 <212> DNA <213> Candida albicans <220> <221> CDS <222> (1) .. (1245) <220> <221> gene <222> (1)..(1245) <223> gene CaIR012 <400> 1 atg tca cac caa caa gaa gac gtc gta gac gat act caa gaa gaa tat Met Ser His Gln Glu Asp Val Val Asp Asp Thr Gln Glu Glu Tyr 5 1 atc aat gtt aat gaa gtg gct gag gaa gtt gca gat gat gat caa gcg Ile Asn Val Asn Glu Val Ala Glu Glu Val Ala Asp Asp Asp Gln Ala 25 20 Pro Pro Asp Glu Glu Asp Glu Glu Met Glu Leu Asp Asp Glu His Glu 45 35 act tta gaa att gac atg tcc aac aat tca tgg act tat ttt gat aaa Thr Leu Glu Ile Asp Met Ser Asn Asn Ser Trp Thr Tyr Phe Asp Lys

55

cat acc gat agt ata ttt act att ttt tca cat cct aaa ttg cca atg

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gtc	acg	gtc	aaa	gta	cat	ggt	aat	ctt	gṛg	gcc	att	ggt	ggc	aga	gat	816

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Thr	Leu 50	Glu	Ile	Asp	Met	Ser 55	Asn	Asn	Ser	Trp	Thr 60	Tyr	Phe	Asp	Lys
lis 65	Thr	Asp	Ser	Ile	Phe 70	Thr	Ile	Phe	Ser	His 75	Pro	Lys	Leu	Pro	Met 80
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Thr	Gln	Pro	Pro 100	Arg	Phe	Val	Gly	Glu 105	Ile	Thr	Gly	His	Lys 110	Glu	Ser
Val	Ile	Ser 115	Gly	Gly	Phe	Thr	Ala 120	Asp	Gly	Lys	Phe	Val 125	Val	Thr	Ala
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Phe	Ile 210	Gln	Gly	Lys	Asp	Glu 215	Asn	Asp	Leu	Thr	Leu 220	Val	Ser	Ile	Ser
Glu 225	Asp	Gly	Thr	Val	Val 230	Asn	Trp	Asn	Cys	Phe 235	Thr	Gly	Gln	Val	Asn 240
Tyr	Lys	Leu	Gln	Pro 245		Asp	Asp	Phe	Lys 250		Val	Glu	Ser	Pro 255	Trp
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Leu	Lys 290	Thr	Leu	Asp	Asn	Val 295	Asp	Asp	Ile	Ala	Glu 300	Leu	Ser	Ile	Glu
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### SEQUENCE LISTING

<110> Hoechst Marion Roussel <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES <130> SEQID07 <140> <141> <160> 2 <170> PatentIn Ver. 2.1 <210> 1 <211> 5544 <212> DNA <213> Candida albicans <220> <221> CDS <222> (1)..(5541) <220> <221> gene <222> (1)..(5541) <223> Gene CaJLO39 <400> 1 atg agt ggc ata ttt aat tgg tcg ctg gat gtg ttt gcc gat att tat Met Ser Gly Ile Phe Asn Trp Ser Leu Asp Val Phe Ala Asp Ile Tyr 15 10 aac acc ctc aag ttt gag tcc aat ata gat ttg gat aca atc gac ttc Asn Thr Leu Lys Phe Glu Ser Asn Ile Asp Leu Asp Thr Ile Asp Phe 25 20 acc agc atc aag aat gat ctt gca aat gtt ttg att aca cca gtc cct Thr Ser Ile Lys Asn Asp Leu Ala Asn Val Leu Ile Thr Pro Val Pro 40 ctg gat caa tca cgt agc aaa ctt gga gac gca tca aaa cca gtg gcg Leu Asp Gln Ser Arg Ser Lys Leu Gly Asp Ala Ser Lys Pro Val Ala 55 50

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240

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														tat Tyr		384
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act Thr 145	aac Asn	aac Asn	aac Asn	caa Gln	gtg Val 150	ttg Leu	ttt Phe	gac Asp	aat Asn	att Ile 155	ttg Leu	aaa Lys	agt Ser	ttc Phe	agc Ser 160	480
aag Lys	att Ile	tat Tyr	act Thr	ttg Leu 165	agt Ser	ggt Gly	aaa Lys	tta Leu	aat Asn 170	gac Asp	atg Met	att Ile	gac Asp	aag Lys 175	caa Gln	528
aaa Lys	gtt Val	acc Thr	ġgc Gly 180	gac Asp	atc Ile	aac Asn	aat Asn	ctt Leu 185	gca Ala	ttt Phe	atc Ile	aat Asn	tgt Cys 190	atc Ile	aat Asn	576
tat Tyr	tcc Ser	aga Arg 195	agt Ser	cag Gln	ttg Leu	ttt Phe	aat Asn 200	gca Ala	cac	gag Glu	tta Leu	ttg Leu 205	gga Gly	caa Gln	gtt Val	624
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3 2 F	<b>~</b> 22	agt	gac	gaa	αaα	gaa	gaa	gaa	gaa	qaa	agc	gac	agc	gac	gaa	1440
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465					470					4/3				•	400	
gat	tct	tcg	acc	cca	aaa	aac	aaa	gaa	aaa	tca	gct	999	tta	gac	ctt	1488
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Ala	Tvr	Thr	Tyr	Asn	Asn	Arg	Pro	Glu	Leu	Cys	Ala	Leu	Phe	Trp	Gly	
	- ]	515	-1-			_	520					525				
		515					320									
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Ile	Leu	vaı		ASI	ASI	ASII	ASII		361	****	*** 9	_,_	590			
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Ser	Lys	Val	Ser	Val	Asp	Ser	Leu	Tyr	Asp	Ser	Leu	Lys	Tyr	Tyr	Ile	
	4	595			-		600					605				
		333														
			aat				<b>~</b> 33	C 2 2	gat	tta	aat	acc	caa	tta	atq	1872
gac	tct	tta	aac	gaa	agc		gaa	Caa	940	Tan	200	712	Gln	T.e.11	Met	
Asp	Ser	Leu	Asn	Glu	Ser	Phe	GIu	GIN	Asp	Pen		Ala	GIII	neu	1100	
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Len	Asn	Gln	Lys	Lys	Gln	Asp	Phe	Leu	Phe	Ser	Thr	Thr	Thr	Ser	Lys	
625			- 1 -	4 -	630	•				635					640	
U Z 3																
					<u> </u>		<b></b> -		242	2++	att	ata	gag	tta	qcc	1968
cag	gac	ctt	gat	gat	tct	ggc	gag	aac	aya	all	911	aca	コ〜コ	3	5	

Gln	Asp	Leu	Asp	Asp 645	Ser	Gly	Glu	Asn	Arg 650	Ile	Val	Ile	Glu	Leu 655	Ala	
gag Glu	gat Asp	tca Ser	ctt Leu 660	gtc Val	ctc Leu	att Ile	tca Ser	999 Gly 665	ttt Phe	att Ile	caa Gln	tta Leu	ctt Leu 670	tct Ser	gca Ala	2016
														atc		2064
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att Ile	caa Gln	agc Ser	aca Thr	aac Asn 725	aac Asn	ccc Pro	aaa Lys	ttt Phe	att Ile 730	gat Asp	ttg Leu	cca Pro	aat Asn	gtt Val 735	ttc Phe	2208
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Ile	Trp 770													agt Ser		2352
cca Pro 785	gaa	gac Asp	aag Lys	aaa Lys	gat Asp 790	gat Asp	gct Ala	ttt Phe	aga Arg	cat His 795	att Ile	ąag Lys	aga Arg	aag Lys	tat Tyr 800	2400
aac Asn	agt Ser	aag Lys	aaa Lys	aat Asn 805	gtt Val	ccc	atc Ile	aat Asn	caa Gln 810	gca Ala	ttt Phe	tca Ser	aca Thr	aac Asn 815	cta Leu	2448
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Leu	Thr	Pro 835	туr	Ala	Asp	Ser	Asn 840	Glu	Ala	Phe	Thr	Lys 845	Tyr	Ser	Leu	
tta	tat	cct	tat	gac	tta	qqa	tta	999	tat	aga	ttc	aac	aac	caa	ctt	2592
					Leu											
	850		·.	-		855					860			-		
					att											2640
Gly	Ile	Trp	Pro	Tyr	Ile	Glu	Phe	Leu	Met		Asn	Val	Phe	Ala		
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				~at	aat	222	caa	aat	agg	atc	aac	tta	caa	ctt	aat	2688
					Asn											
Ser	GIY	1111	116	885	ng	275			890		•			895		
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			900					905					910			
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Leu	Ile	Asp	Val	Ala	Pro	Lys		Ile	Arg	Asp	Leu		Asn	Phe	Asn	
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					Leu											
_	930	Pne	Asp	ser	пец	935	110	Gry	***	<b>U</b>	940		•			
	, 930															
ttt	qtc	aaa	ttg	cat	cat	tca	gtt	gct	gtg	att	aac	tat	cta	ttt	gaa	2880
					His											
945					950					955					960	
					gcc											2928
Asn	Arg	Thr	Phe		Ala	Leu	Phe	Lys		Val	Asn	IIe	GIY		Asp	
				965					970					975		
·· <del>·····</del>				+	ġgt	~==	tca	aca	gaa	tta	ata	tca	cat	acc	ctt	2976
Cor	grg	aat	gaa	Car	Gly	Glu	Ser	Δla	Glu	Leu	Val	Ser	His	Ala	Leu	
Ser	vai	ASII	980	361	Gry	024	502	985					990			
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					Leu											
-		995				:	1000				:	1005				
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					cga											3072
		Pro	Ile	Leu	Arg		Lys	Asp	Thr			Gln	Leu	Hls	Arg	
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999	aca	900	act	. 333	all	220			3	-5-		ن - ر ان - ر		-		

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Glv	Thr	Ala	Ile	Gly	Ile	Gly	Thr	Ser	Met	Ser	Leu	Ala	Leu	Ala	Thr	
1025					1030					035					040	
1023																
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cct	aga	acc	ata	ttt	gat	tgt	ata	tac	tat	cca	aag	aat	ctg	gga	aca	3168
Pro	Ara	Thr	Ile	Phe	Asp	Cys	Ile	Tyr	Tyr	Pro	Lys	Asn	Leu	Gly	Thr	
	5			1045	_	•			050				1	.055		
			J	1043				-	• • •							
																2216
cat	ggt	gtt	gct	gat	ttt	tac	gaa	gtg	ata	ttg	ttc	cac	tta	tçt	gca	3216
Hic	Glv	Val	Ala	Asp	Phe	Tyr	Glu	Val	Ile	Leu	Phe	His	Leu	Ser	Ala	
	01,					•		1065					070			
			1060													
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- Val	Val	Gln	Phe	Ala	Leu	Tyr	Val	Ser	Cys	Glu	Asn	Thr	Ile	Ser	Asn	
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Luc	- 11-	Tle	Ser	Tle	Leu	Lvs	Gly	Val	Ser	Gln	Ser	Lys	Phe	Phe	Val	
			002				•				100					
1	.090					L095				•						
acc	aga	gtt	tca	agc	tct	gct	gat	ccc	tta	ctc	aac	aac	gat	aga	ttg	3360
mb.~	7~~	1721	Car.	Ser	Ser	Ala	Asp	Pro	Leu	Leu	Asn	Asn	Asp	Arg	Leu	
		vai	SCI							1115				1	120	
1105	<b>;</b>				1110					.113				_	· <b>-</b> - +	
att	acc	aca	ttt	gaa	aac	atc	gac	gag	tca	ata	aaa	atc	aag	ttt	gct	3408
		mb	Dha	G1	) Nan	Tle	Aen	Glu	Ser	Ile	Lvs	Ile	Lys	Phe	Ala	
He	Thr	Thr			ASII	110	vab				-4-		•	1135		
				1125				1	130				-			
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200	71-	3	Tue	Dhe	Glu	Glu	T.eu	Glu	Asp	Ser	Leu	Asn	Met	Lys	Tyr	
Pne	TTE			PILE	Giu	GIU						-	1150	-	_	
		:	1140				•	1145					1130			
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949	-1		3	nh-	17-1	Tou	Gly	Asn	ī.eu	Asn	Gln	Phe	asp	Gly	Lys	
GIu	11e	Leu	Asp	Pne	. var			ns		••••				•	-	
		1155					1160					1165				
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gug	gcc			-1	•••	55-	7	Tau	Clv	There	Lve	va1	LVS	Glv	Asp	
Val	Ala	Thr	Thr	Ala	HIS	Pne	Leu	Leu	GIY			741	-,-	,		
:	1170					1175					1180					
		~~~	++~	ata	CaG	aca	aac	gat	caa	aac	aca	tta	cta	aaa	tct	3600
aca	tta	gac	LLG	gta	cag			3	01-	3.00	Th.	Lau	1.011	Lve	Ser	
Thr	Leu	Asp	Leu	Val	Gln	Thr	Asn	Asp			1111	Deu.	пси	Ly S		
118	5				1190				:	1195					1200	
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ttc	τta	aat	aca	ttg	agc	عدد	ayı	-	900			0	01	T1.	700	
Phe	Leu	Asn	Thr	Leu	Ser	Ile	Ser	Leu	qaA	Leu	116	ser			vah	
				1205					1210	•				1215		
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tac	aat	aat	ggt	. aac	aac	cat	all	acc	500	500	220					

Tyr Asn Asn Gly Asn Asn His Ile Ile Asp Val Gly Pro Ala Lys Leu	
1220 1225 1230	
tog tog ttg att tta dag att ott atd aag ttg tgd daa gat oda att	3744
Ser Ser Leu Ile Leu Gln Ile Leu Ile Lys Leu Cys Gln Asp Pro Ile	
1245	
1235 1240 1243	
tog tog toa ata aca ttg aat caa tta ogt gaa tat gaa gaa ttg ttt	3792
Ser Ser Ser Ile Thr Leu Asn Gln Leu Arg Glu Tyr Glu Glu Leu Phe	
1250 1255 1260	
gaa aaa ttg gtt aac tgt caa cct aaa ctt gat ttg aat acc gtt tgg	3840
Glu Lys Leu Val Asn Cys Gln Pro Lys Leu Asp Leu Asn Thr Val Trp	
1265 1270 1275 1280	
	2000
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Cys Gly Asn Gln Phe Asp Gly Asp Leu Gln Ile Asp Ala Ser Asn Val	
1285 1290 1295	
The set too tet too tet att and	3936
ttt gtt gac aac caa gca age acc cag get tte ttt tee ttt att aac	3,300
Phe Val Asp Asn Gln Ala Ser Thr Gln Ala Phe Phe Ser Phe Ile Asn	
1300 1305 1310	
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Gln Arg Asn Leu Ile Leu Gln Tyr Leu Ser Leu Glu Phe His Ser Val	
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1313	
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Lys Ser Arg Thr Lys Arg Glu Tyr Tyr Ser Lys Val Leu Thr Asn Asp	
· 1330 1340	
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Lys Glu Phe Val Asn Arg Thr Pro Lys Val Leu Thr Phe Leu Asn Ile	
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··· <b>········</b>	
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Leu Asn Tyr Ser Phe Lys Asn Phe Glu Val Gln Lys Tyr Glu Trp Leu	
1365 1370 1375	
The same and got G22	4176
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Asp Gln Lys Phe Asn Val Ser Leu Leu Leu Ala Glu Val Asn Ala Gln	
1380 1385 1390	
aag aat ggt aca tta gat ttt tct gtt tta aca aag gtt ttc cgt ctt	4224
Lys Asn Gly Thr Leu Asp Phe Ser Val Leu Thr Lys Val Phe Arg Leu	
1405	
1395 1400 1403	
ttg tgc caa acg tca aac tta ata aca ccc gag tca aag caa ttg ttt	4272
ceg ego can acg, con and our arm arm a b b	

										,	_			_		
Leu	Cys	Gln	Thr	Ser	Asn	Leu	Ile	Thr	Pro	Glu	Ser	Lys	Gin	Leu	Phe	
1	410				1	415				1	420					
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gcc	gaa	gaa	att	atg	gcc	gaa	gga	ayı.	-	-1-	~	340	Db -	y-1	mb	
Ala	Glu	Glu	Ile	Met	Val	Glu	Gly	Ser	Lys	11e	Ser	Asp	Pne	vaı	Thr	
1425	,			1	L <b>4</b> 30				1	435				1	440	
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aag	tac	ctg	grg	teg	acc	gac	-	-	gac	9-9	- Cu	7.00	Tues	0.40	Ton	
Lys	Tyr	Leu	Val	Ser	Thr	Asp	Leu	Lys	Asp	vaı	GIN	ьeu			Leu	
			1	1445				1	450				1	L <b>4</b> 55		
		+ ~ ~	+~+	<b>~</b> 2 2	ttg	ata	gag	att	tta	att	act	qac	aqt	gga	atc	4416
cat	LCa	Lgg	- cgc	caa	-	-1	949	71.	7	17-1	The	7	Car	Gly	Tle	
His	Ser	Trp	Cys	GIn	Leu	шe	GIU	He	Leu	val	IIII			Gry	Ile.	
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Asn	Ser	Leu	Asn	Phe	Ile	Leu	GIU	vai	Leu	GIN			116	PIO	Lys	
	1	1475					480				1	1485				
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1	490				1	L495				1	1500					
+ 4 3	<b>+</b> + -	tat	att	++2	ttg	+++	gat	ctt	tat	gat	caq	ctq	act	ctt	gcg	4560
tea	LLA	Lyc	gcc	-	-	21-	340		Museu	2	Cln	Len	ጥኮሎ	T.em	Δla	
Ser	Leu	Cys	Val	Leu	Leu	Phe	Asp	Leu			GIII	Ten	1111			
1505	5			:	1510				1	1515				]	L520	
~~~	202	222	aat	o a a	gat	+++	gca	cĖt	gga	att	qaq	aqa	ttg	atc	ccc	4608
yac	aya	-	990	944	3	56-	210	Lou	014	Tla	Glu	Ara	Len	Tle	Pro	•
Asp	Arg	Lys	GIA	GIu	Asp	Pne	AIA			116	GIU	Arg			110	
			:	1525				1	1530					1535		
++=	+++	cag	act	tat	att	gca	aat	att	ctt	aat	tct	aac	tca	aca	CCC	4656
- LCa	-1	-1		290	Ile	31-	01	т10	T 017	Nen	Cor	) en	Ser	Thr	Pro	
Leu	Phe	GIn	Thr	Cys	TTE	Ala			neu	VPII	JCI					
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250		3	00	3	Leu	Three	Val.	Val	Glv	Asn	Lvs	Phe	Leu	Leu	Lvs	
ser			ser	Asp	neu			V41	O. y	*****					-1-	
	:	1555					1560					1565				
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0	Dha	27	N	21	Ser	Dhe	Len	Tare	Gln	Val	Met	His	Ile	Ile	Lys	
		GIU	ar 9	GIU			ست س	-,5							•	
	1570					1575				•	1580					
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Car		Acn	[.ve	Live	Phe	Pne										
	Val	Asp	Lys			Pne	Gin					•				
Ser 158	Val	Asp	Lys		Phe 1590	Pne	GIII			1595		•			1600	
158	Val 5				1590					1595					1600	
158	Val 5									1595					1600	4848

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Ser	Glu	Gly	Pro	Ser	Arg	Ile	Thr	Ser	Thr	Leu	Phe	Leu	Glu	Ser	Leu	
		-		1605		•			1610					1615		
			, -													
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Val	His	Leu	Gly	Thr	Leu	Val	Lys	Val	Asp	Pne	TTE			Ala	Leu	
		1	620					1625				1	1630			
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116			A311	A.Lu			1640			5		L <b>64</b> 5	•			
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Asp	Ala	Met	Ile	Lys	Leu	Cys	Gln	Glu	Lys	Asn	Ser	Gly	Val	Thr	Leu	
1	.650				1	1655				3	L660					
~a+	a 2 t	++-	a+a	+++	gac	tta	ato	gca	ttc	aaa	σca	acq	cta	tat	ttt	5040
Asp	His	Phe	He	Pne	Asp	Leu	Met	Ald			AIA	TIII	пси	Tyr		
1665	;			1	1670				3	L675				1	1680	
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ttt	qtt	aga	gtg	gcc	aaa	tcg	aaa	aac	ggg	gca	ttg	cag	ttg	att	caa	5088
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- 110	• • • •	*** 9		L685	-,-				1690					1695		
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																E126
														att		5136
Asn	Glu	Leu	Phe	Ser	Ile	Leu	His	Gln	Ser	Lys	Phe	Leu	Gln	Ile	Asp	
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Pro	Asp	He	GIÀ	Leu	Ser			TIE	GIU	GIU			nap	His	בעם	
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Thr	Val	Asn	Val	Asn	Val	Leu	Leu	Asp	Thr	Pro	Leu	Ser	Ile	Thr	Asp	
	730					1735		-			L740					
•	.,50				•											
~~25,													*			5280
ttg	gtg	gat	cca	tac	aag	ttg	cga	agt	gaa	aac	act	ata	LCa	tat		5200
Leu	Val	Asp	Pro	Tyr	Lys	Leu	Arg	Ser	Glu	Asn	Thr	Ile	Ser	Tyr	Phe	
1745	;			1	1750				1	L755				1	1760	
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Glu	Pne	Leu			116	Pne	GIII			1111	****	141				
			1	1765			•	7	1770					1775		
atg	gga	cca	aat	tat	caa	cct	gca	att	att	caa	act	aga	gaa	ctt	atg	5376
														Leu		
	- 4		1780	•				1785					1790			
		•														
								~~-		n +- ~	997	202	ast	ttc	tta	5424
aag	agt	gta	aat	cga	ttg	gtg	gta	ggt	gtt	atg	aaa	aga	yat	ttc	LLY	3424

5472

5520

Lys Ser Val Asn Arg Leu Val Val Gly Val Met Lys Arg Asp Phe Leu 1795 1800 1805

gta gag acc aaa caa att ggt caa ggg ttg tac aag gaa gag agt cac Val Glu Thr Lys Gln Ile Gly Gln Gly Leu Tyr Lys Glu Glu Ser His 1810 1815 1820

gag ttg gta tcg ttg aaa gaa ttg gtg aag ttg ttt att ttg att gat Glu Leu Val Ser Leu Lys Glu Leu Val Lys Leu Phe Ile Leu Ile Asp 1825 1830 1835 1840

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Ser Leu Ala His Tyr Ser Val
1845

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1 5 10 15

Asn Thr Leu Lys Phe Glu Ser Asn Ile Asp Leu Asp Thr Ile Asp Phe 20 25 30

Thr Ser Ile Lys Asn Asp Leu Ala Asn Val Leu Ile Thr Pro Val Pro

Leu Asp Gln Ser Arg Ser Lys Leu Gly Asp Ala Ser Lys Pro Val Ala
50 55 60

Leu Pro Ser Gly Asp Glu Val Lys Leu Asn Gln Ala Ser Ile Glu Ile
65 70 75 80

Thr Gly Val Leu Ser Asn Glu Leu Asp Leu Asp Glu Leu Asn Thr Ala 85 90 95

Glu Leu Leu Tyr Asn Ala Ser Asp Leu Ser Tyr Lys Lys Gly Thr Ser 100 105 110

Ile Gly Asp Ser Ala Arg Leu Ala Tyr Tyr Leu Arg Ala His Tyr Ile 115 120 125

Leu Asn Ile Val Gly Tyr Leu Val Ser His Lys Arg Leu Asp Ile Ile 130 135 140

Thr 145	Asn	Asn	Asn	Gln	Val 150	Leu	Phe	Asp	Asn	11e 155	Leu	Lys	Ser	Phe	Ser 160
Lys	Ile	Tyr	Thr	Leu 165	Ser	Gly	Lys	Leu	Asn 170	Asp	Met	Ile	Asp	Lys 175	Gln
Lys	Val	Thr	Gly 180	Asp	Ile	Asn	Asn	Leu 185	Ala	Phe	Ile	Asn	Суs 190	Ile	Asn
Tyr	Ser	Arg 195	Ser	Gln	Leu	Phe	Asn 200	Ala	His	Glu	Leu	Leu 205	Gly	Gln	Val
Val	Phe 210	Gly	Leu	Ala	Asp	Asn 215	Tyr	Tyr	Glu	Ser	Tyr 220	Gly	Thr	Leu	Asn
Asn 225	Tyr	Asn	Ser	Leu	Val 230	Glu	Phe	Ile	Leu	Lys 235	Asn	Ile	Ser	Asp	Glu 240
Asp	Val	Phe	Val	Ile 245	His	Phe	Leu	Pro	Ser 250	Thr	Leu	Gln	Leu	Phe 255	Lys
Lys	Leu	Leu	Gln 260	Leu	Gly	Glu	Glu	Ser 265	Leu	Val	Asp	Gln	Phe 270	Tyr	Lys
Thr	Ile	Thr 275	Ser	Ser	Ile	Leu	Lys 280	Asp	Tyr	Glu	Ala	Asn 285	Asn	Phe	Ser
Lys	Ser 290	Glu	Asp	Ile	Asp	Leu 295	Ser	Lys	Ser	Lys	Leu 300	Ser	Gly	Phe	Glu
11e 305	Val	Thr	Ser	Phe	Ile 310	Phe	Leu	Thr	Glu	Phe 315	Ile	Pro	Trp	Cys	Lys 320
Gln	Leu	Ser	Ser	Arg 325	Thr	Ala	Lys	Tyr	Asp 330	Phe	Lys	Asp	Asp	Ile 335	Leu
Lys	Tyr	Met	Glu 340	Phe	Leu	Ile	Ser	Tyr 345	Gly	Val	Met	Glu	Arg 350	Leu	Leu
Ser	Tyr	Cys 355	Ser	Glu	Thr	Ser	Asn 360	Ala	Lys	Thr	Gln	Gln 365	Val	Tyr	Asp
Trp	Ser 370	Asn	Met	Tyr	Asp	Phe 375	Arg	Ala	Leu	Leu	Gln 380	Lys	Asn	Phe	Pro
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Asn	Ala	Val	Arg	Pro 405	Gly	Tyr	Glu	Asn	Ile 410	Ser	Lys	Leu	Ile	Asp 415	Ile
Ser	Phe	Leu	Thr 420	Leu	Asp	Pro	Ser	Leu 425	Asn	Glu	Thr	Leu	Val 430	Ser	Pro
Phe	Phe	Gln 435	Ser	Phe	Phe	Ser	Val 440	Phe	Ile	Ser	Asn	Ala 445	Ala	Val	Val
Met	Thr 450	Ser	Leu	Arg	Asp	Ser 455	Glu	Glu	Asp	Phe	Val 460	Leu	Ser	Ser	Leu
Asn 465	Glu	Ser	Asp	Glu	Glu 470	Glu	Glu	Glu	Glu	Glu 475	Ser	Asp	Ser	Asp	Glu 480
Asp	Ser	Ser	Thr	Pro 485	Lys	Asn	Lys	Glu	Lys 490	Ser	Ala	Gly	Leu	Asp 495	Leu
Asp	Lys	Ile	Ala 500	Gln	Arg	Ala	Glu	Leu 505	Glu	Arg	Phe	Tyr	Leu 510	Ala	Phe
Ala	Tyr	Thr 515	туr	Asn	Asn	Arg	Pro 520	Glu	Leu	Cys	Ala	Leu 525	Phe	Trp	Gly
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Ala 545	Asn	Asn	Thr	Ser	Pro 550	Leu	Ile	Thr	Ala	Thr 555	Phe	Cys	Leu	Leu	Leu 560
Gly	Ser	Leu	Ala	Ser 565	Ala	Gly	Ala	Glu	Ala 570	Thr	Ser	Arg	Ile	Trp 575	Glu
Ile	Leu	Val	His 580	Asn	Asn	Asn	Asn	Ala 585	Ser	Thr	Arg	Lys	Asn 590	Asp	Phe
Ser	Lys	Val 595	Ser	Val	Asp	Ser	Leu 600	Tyr	Asp	Ser	Leu	Lys 605	Tyr	Tyr	Ile
Asp	Ser 610	Leu	Asn	Glu	Ser	Phe 615	Glu	Gln	Asp	Leu	Asn 620	Ala	Gln	Leu	Met
Leu 625	Asn	Gln	Lys	Lys	Gln 630	Asp	Phe	Leu	Phe	Ser 635	Thr	Thr	Thr	Ser	Lys 640
Gln	Asp	Leu	Asp	Asp 645	Ser	Gly	Glu	Asn	Arg 650	Ile	Val	Ile	Glu	Leu 655	Ala

Glu	Asp	Ser	Leu 660	Val	Leu	Ile	Ser	Gly 665	Phe	Ile	Gln	Leu	Leu 670	Ser	Ala
Ile	Val	Lys 675	Asn	Leu	Asn	Thr	Lys 680	Asn	Glu	Arg	Ser	Lys 685	Glu	Ile	Lys
Ser	Val 690	Val	Tyr	Thr	Arg	Phe 695	Ser	Pro	Ile	Ile	Lys 700	Gly	Phe	Leu	Lys
Phe 705	Asp	Asn	Leu	Ile	Asn 710	Gly	Ser	Arg	Phe	Leu 715	Gln	Val	Asp	Ala	Ser 720
Ile	Gln	Ser	Thr	Asn 725	Asn	Pro	Lys	Phe	Ile 730	Asp	Leu	Pro	Asn	Val 735	Phe
Val	Ser	Asp	Asp 740	Ser	Arg	Ile	Ile	Leu 745	Thr	Asn	Leu	Ile	Leu 750	Thr	Phe
Leu	Gly	Asp 755	Phe	Val	Thr	Asn	Asp 760	Ser	Asp	Pro	Tyr	11e 765	Arg	Tyr	Glu
Ile	Trp 770	Arg	Leu	Val	Asp	Arg 775	Trp	Met	Tyr	Gln	Gly 780	Leu	His	Ser	Leu
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Asn	Ser	Lys	Lys	Asn 805	Val	Pro	Ile	Asn	Gln 810	Ala	Phe	Ser	Thr	Asn 815	Leu
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Leu	Thr	Pro 835	Tyr	Ala	Asp	Ser	Asn 840	Glu	Ala	Phe	Thr	Lys 845	Tyr	Ser	Leu
Leu	Tyr 850	Pro	Суз	Asp	Leu	Gly 855	Leu	Gly	Tyr	Arg	Phe 860	Asn	Asn	Gln	Leu
Gly 865	Ile	Trp	Pro	Tyr	11e 870	Glu	Phe	Leu	Met	Gln 875	Asn	Val	Phe	Ala	Asn 880
Ser	Gly	Thr	Ile	Ala 885		Lys	Arg	Asp	Arg 890	Val	Asn	Leu	Gln	Leu 895	Asn
Leu	Leu	Glu	Leu 900		Ser	Asn	Ala	Leu 905	Gln	Gly	Val	Asp	Trp 910	Lys	Phe

- Leu Ile Asp Val Ala Pro Lys Ile Ile Arg Asp Leu Lys Asn Phe Asn 915 920 925
- Gly Ile Phe Asp Ser Leu Ile Pro Gly Val Gln Leu Asp Phe Glu Val 930 935 940
- Phe Val Lys Leu His His Ser Val Ala Val Ile Asn Tyr Leu Phe Glu 945 950 955 960
- Asn Arg Thr Phe Ser Ala Leu Phe Lys Leu Val Asn Ile Gly Val Asp 965 970 975
- Ser Val Asn Glu Ser Gly Glu Ser Ala Glu Leu Val Ser His Ala Leu 980 985 990
- Gly Leu Ile Asn Ser Leu Leu Arg Val Gln Asn Ser Phe Ile Asn Lys 995 1000 1005
- Leu Leu Pro Ile Leu Arg Asn Lys Asp Thr Gln Gln Gln Leu His Arg 1010 1015 1020
- Gly Thr Ala Ile Gly Ile Gly Thr Ser Met Ser Leu Ala Leu Ala Thr
  025 1030 1035 1040
- Pro Arg Thr Ile Phe Asp Cys Ile Tyr Tyr Pro Lys Asn Leu Gly Thr
  1045 1050 1055
- His Gly Val Ala Asp Phe Tyr Glu Val Ile Leu Phe His Leu Ser Ala 1060 1065 1070
- Val Val Gln Phe Ala Leu Tyr Val Ser Cys Glu Asn Thr Ile Ser Asn 1075 1080 1085
- Lys Ala Ile Ser Ile Leu Lys Gly Val Ser Gln Ser Lys Phe Phe Val 1090 1095 1100
- Thr Arg Val Ser Ser Ser Ala Asp Pro Leu Leu Asn Asn Asp Arg Leu 105 1110 1115 1120
- Ile Thr Thr Phe Glu Asn Ile Asp Glu Ser Ile Lys Ile Lys Phe Ala 1125 1130 1135
- Phe Ile Asp Lys Phe Glu Glu Leu Glu Asp Ser Leu Asn Met Lys Tyr 1140 1145 1150
- Glu Ile Leu Asp Phe Val Leu Gly Asn Leu Asn Gln Phe Asp Gly Lys
  1155 1160 1165

- Val Ala Thr Thr Ala His Phe Leu Leu Gly Tyr Lys Val Lys Gly Asp 1170 1175 1180
- Thr Leu Asp Leu Val Gln Thr Asn Asp Gln Asn Thr Leu Leu Lys Ser 185 1190 1195 1200
- Phe Leu Asn Thr Leu Ser Ile Ser Leu Asp Leu Ile Ser Glu Ile Asp 1205 1210 1215
- Tyr Asn Asn Gly Asn Asn His Ile Ile Asp Val Gly Pro Ala Lys Leu 1220 1225 1230
- Ser Ser Leu Ile Leu Gln Ile Leu Ile Lys Leu Cys Gln Asp Pro Ile 1235 1240 1245
- Ser Ser Ser Ile Thr Leu Asn Gln Leu Arg Glu Tyr Glu Glu Leu Phe 1250 1255 1260
- Glu Lys Leu Val Asn Cys Gln Pro Lys Leu Asp Leu Asn Thr Val Trp 265 1270 1275 1280
- Cys Gly Asn Gln Phe Asp Gly Asp Leu Gln Ile Asp Ala Ser Asn Val 1285 1290 1295
- Phe Val Asp Asn Gln Ala Ser Thr Gln Ala Phe Phe Ser Phe Ile Asn 1300 1305 1310
- Gln Arg Asn Leu Ile Leu Gln Tyr Leu Ser Leu Glu Phe His Ser Val
- Lys Ser Arg Thr Lys Arg Glu Tyr Tyr Ser Lys Val Leu Thr Asn Asp 1330 1335 1340
- Lyg Glu Phe Val Asn Arg Thr Pro Lys Val Leu Thr Phe Leu Asn Ile
  345 1350 1355 1360
- Leu Asn Tyr Ser Phe Lys Asn Phe Glu Val Gln Lys Tyr Glu Trp Leu 1365 1370 1375
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- Lys Asn Gly Thr Leu Asp Phe Ser Val Leu Thr Lys Val Phe Arg Leu 1395 1400 1405
- Leu Cys Gln Thr Ser Asn Leu Ile Thr Pro Glu Ser Lys Gln Leu Phe 1410 1415 1420

431	83
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- Ala Glu Glu Ile Met Val Glu Gly Ser Lys Ile Ser Asp Phe Val Thr
  425 1430 1435 1440
- Lys Tyr Leu Val Ser Thr Asp Leu Lys Asp Val Gln Leu Lys Cys Leu 1445 1450 1455
- His Ser Trp Cys Gln Leu Ile Glu Ile Leu Val Thr Asp Ser Gly Ile 1460 1465 1470
- Asn Ser Leu Asn Phe Ile Leu Glu Val Leu Gln Val Ile Ile Pro Lys 1475 1480 1485
- Ile Asn Asp Tyr Phe Asp Val Asp Ile Leu Phe Ser Glu Glu Met Val 1490 1495 1500
- Ser Leu Cys Val Leu Leu Phe Asp Leu Tyr Asp Gln Leu Thr Leu Ala 505 1510 1515 1520
- Asp Arg Lys Gly Glu Asp Phe Ala Leu Gly Ile Glu Arg Leu Ile Pro 1525 1530 1535
- Leu Phe Gln Thr Cys Ile Ala Gly Ile Leu Asn Ser Asn Ser Thr Pro 1540 1545 1550
- Ser Leu Arg Ser Asp Leu Tyr Val Val Gly Asn Lys Phe Leu Leu Lys 1555 1560 1565
- Cys Phe Glu Arg Glu Ser Phe Leu Lys Gln Val Met His Ile Ile Lys 1570 1580
- Ser Val Asp Lys Lys Phe Phe Gln Val Ile Cys Asn Asp Ala Ile Tyr 585 1590 1595 1600
- Ser Glu Gly Pro Ser Arg Ile Thr Ser Thr Leu Phe Leu Glu Ser Leu 1605 1610 1615
- Val His Leu Gly Thr Leu Val Lys Val Asp Phe Ile Leu Asn Ala Leu 1620 1625 1630
- Ile Lys Asn Asn Ala Leu Leu Leu Leu Val Arg Ser Val Lys Arg Thr 1635 1640 1645
- Asp Ala Met Ile Lys Leu Cys Gln Glu Lys Asn Ser Gly Val Thr Leu 1650 1655 1660
- Asp His Phe Ile Phe Asp Leu Met Ala Phe Lys Ala Thr Leu Tyr Phe 665 1670 1675 1680

- Phe Val Arg Val Ala Lys Ser Lys Asn Gly Ala Leu Gln Leu Ile Gln 1685 1690 1695
- Asn Glu Leu Phe Ser Ile Leu His Gln Ser Lys Phe Leu Gln Ile Asp 1700 1705 1710
- Pro Asp Ile Gly Leu Ser Leu Arg Ile Glu Glu Val Gln Asp His Lys 1715 1720 1725
- Thr Val Asn Val Asn Val Leu Leu Asp Thr Pro Leu Ser Ile Thr Asp 1730 1735 1740
- Leu Val Asp Pro Tyr Lys Leu Arg Ser Glu Asn Thr Ile Ser Tyr Phe
  745 1750 1755 1760
- Glu Phe Leu Val Pro Ile Phe Gln Leu Leu Thr Thr Val Leu Leu Ser 1765 1770 1775
- Met Gly Pro Asn Tyr Gln Pro Ala Ile Ile Gln Thr Arg Glu Leu Met 1780 1785 1790
- Lys Ser Val Asn Arg Leu Val Val Gly Val Met Lys Arg Asp Phe Leu 1795 1800 1805
- Val Glu Thr Lys Gln Ile Gly Gln Gly Leu Tyr Lys Glu Glu Ser His 1810 1815 1820
- Glu Leu Val Ser Leu Lys Glu Leu Val Lys Leu Phe Ile Leu Ile Asp 825 1830 1835 1840
- Ser Leu Ala His Tyr Ser Val 1845

### SEQUENCE LISTING

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<120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR
 SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES

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WO 00/15838	
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1 5 10 15	
and the second of	gac 96
tog tta cca cca cca cac cca cca aat cct act ggt att gac agt	Jac 90 Asn
Trp Leu Pro Pro Pro His Pro Pro Asn Pro Thr Gly Ile Asp Ser I	
20 25	
ccg gct tta gca cca cat ggt gtt gaa caa gcc caa cag tta gct	gcc 144
Pro Ala Leu Ala Pro His Gly Val Glu Gln Ala Gln Gln Leu Ala	Ala
35 40 45	
	act 192
tat ctt aca tca tta cct aca cat gaa aag cct gaa ttt att att	get 192 Nla
Tyr Leu Thr Ser Leu Pro Thr His Glu Lys Pro Glu Phe Ile Ile	n.a
50 55	
toa cot tit tat ogt tgt ata gaa acg tog aga coc att goo gaa	atg 240

Ser Pro Phe Tyr Arg Cys Ile Glu Thr Ser Arg Pro Ile Ala Glu Met
65 70 75 80

ttg gac ttg aag att gct tta gaa aga gga gtt ggt gaa tgg ttt cgt 288
Leu Asp Leu Lys Ile Ala Leu Glu Arg Gly Val Gly Glu Trp Phe Arg
85 90 95

aaa aat aga gat acc aaa cca gtt ccc ggt gat tac aca caa ttg aga 336 Lys Asn Arg Asp Thr Lys Pro Val Pro Gly Asp Tyr Thr Gln Leu Arg 100 105 110

aca ttt ttc gat aaa tta ttg atc gat gaa gat act tgg cca aga gat 384
Thr Phe Phe Asp Lys Leu Leu Ile Asp Glu Asp Thr Trp Pro Arg Asp
115 120 125

aac tta aat gtt ata cct aat att gaa gga gaa gat tat gat gaa atc 432
Asn Leu Asn Val Ile Pro Asn Ile Glu Gly Glu Asp Tyr Asp Glu Ile
130 135 140

tac gat cgt gcc aaa ttg ttt tgg aaa aag ttt att cct gaa ttt gaa 480 Tyr Asp Arg Ala Lys Leu Phe Trp Lys Lys Phe Ile Pro Glu Phe Glu 145 150 155 160

aag aaa ttc ccc gaa att aaa aat gtg ttg ata gtt aca cat gca gca 528 Lys Lys Phe Pro Glu Ile Lys Asn Val Leu Ile Val Thr His Ala Ala 165 170 175

acg aaa att gct tta gga tca gct tta tta cag tta aaa tca gtt act 576
Thr Lys Ile Ala Leu Gly Ser Ala Leu Leu Gln Leu Lys Ser Val Thr
180 185 190

gat gtt ata gat gat aat caa act gtg tta cgt gct ggt gca tgt tca 624 Asp Val Ile Asp Asp Asn Gln Thr Val Leu Arg Ala Gly Ala Cys Ser 195 200 205

tta tcc aaa ttt gtt aga gat ggc gaa gat aaa acc aat cat act att

Leu Ser Lys Phe Val Arg Asp Gly Glu Asp Lys Thr Asn His Thr Ile

210 215 220

caa tgg aaa att gtc atg aat ggt aat tgt gaa ttc ttg aca cag ggt 720
Gln Trp Lys Ile Val Met Asn Gly Asn Cys Glu Phe Leu Thr Gln Gly
225 230 235 240

gaa gaa atg aac tgg gat ttc cgt cgt ggt gtt gaa gcc ggg tca gct 768
Glu Glu Met Asn Trp Asp Phe Arg Arg Gly Val Glu Ala Gly Ser Ala
245 250 255

gaa gat ata gcg caa aga aag gca gca gca gaa gca gaa gca aaa gca 816

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921

Glu Asp Ile Ala Gln Arg Lys Ala Ala Ala Glu Ala Glu Ala Lys Ala 260 265 270

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Leu Lys Lys Asn Glu Gln Thr Lys Ser Asp Gly Pro Ile Thr Glu Ser
275 280 285

gcc act ggg gca gaa ata gat ggg aat gaa gat gaa ttt gaa gta cgt 912 Ala Thr Gly Ala Glu Ile Asp Gly Asn Glu Asp Glu Phe Glu Val Arg 290 295 300

aaa act tga Lys Thr

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Trp Leu Pro Pro Pro His Pro Pro Asn Pro Thr Gly Ile Asp Ser Asp
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10

Pro Ala Leu Ala Pro His Gly Val Glu Gln Ala Gln Gln Leu Ala Ala 35 40 45

Tyr Leu Thr Ser Leu Pro Thr His Glu Lys Pro Glu Phe Ile Ile Ala 50 55 60

Ser Pro Phe Tyr Arg Cys Ile Glu Thr Ser Arg Pro Ile Ala Glu Met
65 70 75 80

Leu Asp Leu Lys Ile Ala Leu Glu Arg Gly Val Gly Glu Trp Phe Arg

Lys Asn Arg Asp Thr Lys Pro Val Pro Gly Asp Tyr Thr Gln Leu Arg 100 105 110

Thr Phe Phe Asp Lys Leu Leu Ile Asp Glu Asp Thr Trp Pro Arg Asp

Asn Leu Asn Val Ile Pro Asn Ile Glu Gly Glu Asp Tyr Asp Glu Ile 130 135 140

	83
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Tyr 145	Asp	Arg	Ala	Lys	Leu 150	Phe	Trp	Lys	Lys	Phe 155	Ile	Pro	Glu	Phe ·	Glu 160
Lys	Lys	Phe	Pro	Glu 165	Ile	Lys	Asn	Val	Leu 170	Ile	Val	Thr	His	Ala 175	Ala

- Thr Lys Ile Ala Leu Gly Ser Ala Leu Leu Gln Leu Lys Ser Val Thr 180 185 190
- Asp Val Ile Asp Asp Asn Gln Thr Val Leu Arg Ala Gly Ala Cys Ser 195 200 205
- Leu Ser Lys Phe Val Arg Asp Gly Glu Asp Lys Thr Asn His Thr Ile 210 215 220
- Gln Trp Lys Ile Val Met Asn Gly Asn Cys Glu Phe Leu Thr Gln Gly 225 230 235 240
- Glu Glu Met Asn Trp Asp Phe Arg Arg Gly Val Glu Ala Gly Ser Ala 245 250 255
- Glu Asp Ile Ala Gln Arg Lys Ala Ala Ala Glu Ala Glu Ala Lys Ala 260 265 270
- Leu Lys Lys Asn Glu Gln Thr Lys Ser Asp Gly Pro Ile Thr Glu Ser 275 280 285
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Lys Thr

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SEQUENCE LISTING

<110> Hoechst Marion Roussel <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES <130> SEQID10 <140> <141> <160> 2 <170> PatentIn Ver. 2.1 <210> 1 <211> 1454 <212> DNA <213> Artificial Sequence <220> <221> CDS <222> (1)..(1452) / <223> Gene CaOR110 Splice Variant <223> Description of Artificial Sequence: Splice Vaiant atg acg att gaa act att tat atc gca aga cac ggt tat aga tcc aat Met Thr Ile Glu Thr Ile Tyr Ile Ala Arg His Gly Tyr Arg Ser Asn tgg tta cca cca cca cca cca aat cct act ggt att gac agt gac Trp Leu Pro Pro Pro His Pro Pro Asn Pro Thr Gly Ile Asp Ser Asp 30 20 ceg get tta gea cea cat ggt gtt gaa caa gee caa cag tta get gee Pro Ala Leu Ala Pro His Gly Val Glu Gln Ala Gln Gln Leu Ala Ala 40 tat ctt aca tca tta cct aca cat gaa aag cct gaa ttt att att gct 192 Tyr Leu Thr Ser Leu Pro Thr His Glu Lys Pro Glu Phe Ile Ile Ala 50 55 tca cct ttt tat cgt tgt ata gaa acg tcg aga ccc att gcc gaa atg Ser Pro Phe Tyr Arg Cys Ile Glu Thr Ser Arg Pro Ile Ala Glu Met

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Leu	Asp	Leu	Lys	Ile	Ala	Leu	Glu	Arg	Gly	Val	Gly	Glu	Trp	Phe	Arg	
	•		•	85					90					95		
aaa	aat	aga	gat	acc	aaa	cca	gtt	ccc	ggt	gat	tac	aca	caa	ttg	aga	336
Lvs	Asn	Arg	Asp	Thr	Lys	Pro	Val	Pro	Gly	Asp	Tyr	Thr	Gln	Leu	Arg	
-, -		_	100					105					110			
																5
aca	ttt	ttc	gat	aaa	tta	ttg	atc	gat	gaa	gat	act	tgg	cca	aga	gat	384
Thr	Phe	Phe	Asp	Lys	Leu	Leu	lle	Asp	Glu	Asp	Thr	Trp	Pro	Arg	Asp	
		115	-				120					125		•		
									~							
aac	tta	aat	gtt	ata	cct	aat	att	gaa	gga	gaa	gat	tat	gat	gaa	atc	432
Asn	Leu	Asn	Val	Ile	Pro	Asn	Ile	Glu	Gly	Glu	Asp	Tyr	Asp	Glu	Ile	•
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tac	gat	cgt	gcc	aaa	ttg	ttt	tgg	aaa	aag	ttt	att	cct	gaa	ttt	gaa	480
Tyr	Asp	Arg	Ala	Lys	Leu	Phe	Trp	Lys	Lys	Phe	Ile	Pro	Glu	Phe	Glu	
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Lys	Lys	Phe	Pro	Glu	Ile	Lys	Asn	Val	Leu	Ile	Val	Thr	His	Ala	Ala	
•	-			165					170					175		
acg	aaa	att	gct	tta	gga	tca	gct	tta	tta	cag	tta	aaa	tca	gtt	act	576
Thr	Lys	Ile	Ala	Leu	Gly	Ser	Ala	Leu	Leu	Gln	Leu	Lys	Ser	Val	Thr	
	_		180					185					190			
gat	gtt	ata	gat	gat	aat	caa	act	gtg	tta	cgt	gct	ggt	gca	tgt	tca	624
Asp	Val	Ile	Asp	Asp	Asn	Gln	Thr	Val	Leu	Arg	Ala	Gly	Ala	Cys	Ser	
	•	195					200					205				
tta	tcc	aaa	ttt	gtt	aga	gat	ggc	gaa	gat	aaa	acc	aat	cat	act	att	672
Leu	Ser	Lys	Phe	Val	Arg	Asp	Gly	Glu	Asp	Lys	Thr	Asn	His	Thr	Ile	
	210					215					220					
	•															
çaa	tgg	aaa	att	gtc	atg	aat	ggt	aat	tgt	gaa	ttc	ttg	aca	cag	ggt	720
Gln	Trp	Lys	Ile	Val	Met	Asn	Gly	Asn	Cys	Glu	Phe	Leu	Thr	Gln	GIA	
225					230					235					240	
												·				7.0
gaa	gaa	atg	aac	tgg	gat	ttc	cgt	cgt	ggt	gtt	gaa	gcc	999	tça	gct	768
Glu	Glu	Met	Asn	Trp	Asp	Phe	Arg	Arg		Val	Glu	Ala	Gly	Ser	Ala	
				245					250	)				255		
															~~~	816
gaa	gat	ata	gcg	caa	aga	aag	gca	gca	gca	gaa	gca	gaa	gca	aaa	gca	910
<b>~</b> 3	•	T10	. או	015	λ×α	LVG	. Ala	בו ב	. או	Glu	Ala	لللك	АТА	. ∟yS	WIG	

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270 260

ttg aag aaa aat gaa caa acc aaa tcc gat ggt ccc atc act gaa tct Leu Lys Lys Asn Glu Gln Thr Lys Ser Asp Gly Pro Ile Thr Glu Ser 285 280 275 gcc act ggg gca gaa ata gat ggg aat gaa gat gaa ttt gaa aca ttt Ala Thr Gly Ala Glu Ile Asp Gly Asn Glu Asp Glu Phe Glu Thr Phe tat gta acc atc gat ata cct tca att tcg aat aaa atc gac aat gaa Tyr Val Thr Ile Asp Ile Pro Ser Ile Ser Asn Lys Ile Asp Asn Glu 315 310 305 gaa gaa cca cca tca agg aca ggt caa gct cca aaa ttc aaa aac aat 1008 Glu Glu Pro Pro Ser Arg Thr Gly Gln Ala Pro Lys Phe Lys Asn Asn 325 att atc aag cct tca gca caa ctc caa ttt act gat tta aaa gaa gat Ile Ile Lys Pro Ser Ala Gln Leu Gln Phe Thr Asp Leu Lys Glu Asp 345 340 cat cca tta gta aaa ata tcg aac aat act ata tct gct caa ggc tcg His Pro Leu Val Lys Ile Ser Asn Asn Thr Ile Ser Ala Gln Gly Ser 360 tog tog tog tog that ca gog tog aaa aat gga tit aat agt cat act Ser Ser Ser Leu Ser Ala Ser Lys Asn Gly Phe Asn Ser His Thr 380 370 375 cac aat toa gga gto att gat coa toa goa ott ata gat ggg aaa att 1200 His Asn Ser Gly Val Ile Asp Pro Ser Ala Leu Ile Asp Gly Lys Ile 400 390 385 tat cag act gat tgg aat caa tta caa ggt act gaa cta ata ttt gat 1248 Tyr Gln Thr Asp Trp Asn Gln Leu Gln Gly Thr Glu Leu Ile Phe Asp 410 405 gaa aat ggt caa ttt ata ggc aag gtt aag gaa cat ttg act tgc aat 1296 Glu Asn Gly Gln Phe Ile Gly Lys Val Lys Glu His Leu Thr Cys Asn 425 430 420 aat aac aca aaa ttc aca tta aaa aag gca gaa gaa gta gaa caa ctt 1344 Asn Asn Thr Lys Phe Thr Leu Lys Lys Ala Glu Glu Val Glu Gln Leu 440 cgt tea gea gat gat tet ate atg gat ata gat caa gae tea caa gga 1392 Arg Ser Ala Asp Asp Ser Ile Met Asp Ile Asp Gln Asp Ser Gln Gly

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450

455

caa caa cca gct aga agt cag ttc tta aaa aga gca att gtg gct gct 1440
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aga gcc aaa ggt aa Arg Ala Lys Gly 1454

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- <212> PRT
- <213> Artificial Sequence
- <223> Description of Artificial Sequence: Splice Vaiant

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Tyr Leu Thr Ser Leu Pro Thr His Glu Lys Pro Glu Phe Ile Ile Ala 50 55 60

Ser Pro Phe Tyr Arg Cys Ile Glu Thr Ser Arg Pro Ile Ala Glu Met 65 70 75 80

Leu Asp Leu Lys Ile Ala Leu Glu Arg Gly Val Gly Glu Trp Phe Arg 85 90 95

Lym Asn Arg Asp Thr Lys Pro Val Pro Gly Asp Tyr Thr Gln Leu Arg 100 105 110

Thr Phe Phe Asp Lys Leu Leu Ile Asp Glu Asp Thr Trp Pro Arg Asp 115 120 125

Asn Leu Asn Val Ile Pro Asn Ile Glu Gly Glu Asp Tyr Asp Glu Ile 130 135 140

Tyr Asp Arg Ala Lys Leu Phe Trp Lys Lys Phe Ile Pro Glu Phe Glu 145 150 155 160

Lys Lys Phe Pro Glu Ile Lys Asn Val Leu Ile Val Thr His Ala Ala

165

Thr	Lys	Ile	Ala 180	Leu	Gly	Ser	Ala	Leu 185	Leu	Gln	Leu	Lys	Ser 190	Val	Thr
Asp	Val	Ile 195	Asp	Asp	Asn	Gln	Thr 200	Val	Leu	Arg	Ala	Gly 205	Ala	Cys	Ser
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Glu	Glu	Met	Asn	Trp 245	Asp	Phe	Arg	Arg	Gly 250	Val	Glu	Ala	Gly	Ser 255	Ala
Glu	Asp	Ile	Ala 260	Gln	Arg	Lys	Ala	Ala 265	Ala	Glu	Ala	Glu	Ala 270	Lys	Ala
Leu	Lys	Lys 275	Asn	Glu	Gln	Thr	Lys 280	Ser	Asp	Gly	Pro	Ile 285	Thr	Glu	Ser
Ala	Thr 290	Gly	Ala	Glu	Ile	Asp 295	Gly	Asn	Glu	Asp	Glu 300	Phe	Glu	Thr	Phe
Tyr 305	Val	Thr	Ile	Asp	Ile 310	Pro	Ser	Ile	Ser	Asn 315	Lys	Ile	Asp	Asn	Glu 320
Glu	Glu	Pro	Pro	Ser 325	Arg	Thr	Gly	Gln	Ala 330	Pro	Lys	Phe	Lys	Asn 335	Asn
Ile	Ile	Lys	Pro 340	Ser	Ala	Gln	Leu	Gln 345	Phe	Thr	Asp	Leu	Lys 350	Glu	Asp
нів	Pro	Leu 355		Lys	Ile	Ser	Asn 360	Asn	Thr	Ile	Ser	Ala 365	Gln	Gly	Ser
Ser	Ser 370		Ser	Leu	Ser	Ala 375		Lys	Asn	Gly	Phe 380	Asn	Ser	His	Thr
His 385	Asn	Ser	Gly	Val	Ile 390	Asp	Pro	Ser	Ala	Leu 395		Asp	Gly	Lys	Ile 400
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Glu	Asn	Gly	Gln	Phe	lle	Gly	Lys	Val	Lys	Glu	His	Leu	Thr	Cys	Asn

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Asn Asn Thr Lys Phe Thr Leu Lys Lys Ala Glu Glu Val Glu Gln Leu 435 440 445

Arg Ser Ala Asp Asp Ser Ile Met Asp Ile Asp Gln Asp Ser Gln Gly
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Gln Gln Pro Ala Arg Ser Gln Phe Leu Lys Arg Ala Ile Val Ala Ala 465 470 475 480

Arg Ala Lys Gly

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#### SEQUENCE LISTING

<110> Hoechst Marion Roussel <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES <130> SEQID11 <140> <141> <160> 2 <170> PatentIn Ver. 2.1 <210> 1 . <211> 2877 <212> DNA <213> Candida albicans · <220> <221> CDS <222> (1)..(2874) <220> <221> gene <222> (1)..(2874) <223> Gene CaMR212 <400> 1 atg aat ttg ttt caa cat aaa cat caa aaa tta ata tta caa tgt tat Met Asn Leu Phe Gln His Lys His Gln Lys Leu Ile Leu Gln Cys Tyr 10 cct gct ggg aaa gca gtg gac aaa aaa ccc aac tcg tcc gag tta agt Pro Ala Gly Lys Ala Val Asp Lys Lys Pro Asn Ser Ser Glu Leu Ser 20 tat tta tta tac tat gca tcc act cgt aga gtc aaa tta gaa aag gtg Tyr Leu Leu Tyr Tyr Ala Ser Thr Arg Arg Val Lys Leu Glu Lys Val 40 35 att aat tit tig aaa gat aaa act cat cat gat git ggt aga aac cgt Ile Asn Phe Leu Lys Asp Lys Thr His His Asp Val Gly Arg Asn Arg

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					aac Asn											336
					gtt Val											384
					ttt Phe											432
					aag Lys 150											480
					atg Met											528
					ggt Gly											576
					att Ile											624
					ctc Leu											672
					aaa Lys 230											720
					gat Asp											768
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Ala	Phe	Ser	Ser 260	Met	Lys	Ser	Phe	Phe 265		Thr	Asn	Ala	Ala 270	Ser	Gln	
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											tta Leu 300					912
											acc Thr					960
											tcc Ser					1008
											ctt Leu					1056
											caa Gln					1104
											ttg Leu 380					1152
											att Ile					1200
											tcg Ser					1248
											ttt Phe					1296
											ttg Leu					1344
ttg	gat	aac	att	tcg	aag	att	ttt	tta	att	ttg	aag	aat	aaa	tca	agc	1392

Leu	Asp 450	Asn	Ile	Ser	Lys	Ile 455	Phe	Leu	Ile	Leu	Lys 460	Asn	Lys	Ser	Ser	
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Ser	Ile	Asn	Arg	Asn	His	Val	Asn	Leu	Glu	His	Trp	Asp	Ile	Ser	Leu	
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Ile	Ser	Thr	Thr	Gln	Leu	Ile	Asn	Ile	Gln	Ala	Arg	Tyr	Leu	Lys	Val	
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	_				aat											1584
Phe	Asp	Glu	Phe	Leu	Asn	Asn	Glu	Leu	Ala	Val	Gly	Asn	Ser	Lys	Lys	
		515					520					525				
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_	_	-			aac											1680
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545					550					555					560	
																1728
					gcc											1/20
Phe	Lys	ГÀЗ	Pro		Ala	Asn	GIN	туг		Int	ASII	GIII	GIII	575	FIIC	
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					Met											
116	Ser	птэ	580	neu	Mec	- 7 -	110	585	2,0				590	-1-		
			300					303								
+ C C	ccc	aac	aca	caa	tca	ata	tta	ctt	tta	att	act	att	tta	aaa	gat	1824
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501		595	****	<b></b>			600					605 <sup>.</sup>		•	-	
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			ttg													2016
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			aaa													2064
Lys	Ser	Ser	Lys	Leu	Phe	Lys	Gln	Ile	Leu	Asp	Ala	Val	Glu	Tyr	Arg	
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aaa	atg	caa	aag	בננ	cgg	gue.	cac	990		940	_	2	D	2	3	
Lys	Met	Gln	Lys	Phe	Trp	Val	His	GIA	Ile	Asp	Pro	Ser	Pro	ser	Asp	
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Leu	Glu	Asn	Thr	Lys	Gly	Asp	Arg	Thr	116		1111	ASP	AIG	Wali		
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3	Th. 4 = 4	710	Ala	710	۸۳۵	Tle	Lvs	Pro	Glu	Asn	Ile	Glu	Glu	Phe	Ala	
ASII	TAT	TIE	AIG		ur 9		2,0		730					735		
				725					/30					,,,,		
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		755			•		760					765				
		, , ,														
	•							- 4					~a+	<b></b>	<b>a</b> aa	2352
aat	aat	gat	tct	aga	aac	aca	aat	atg	aca	geg	ata	acy	gat	caa	994	2372
Asn	Asn	Asp	Ser	Arg	Asn	Thr	Asn	Met	Thr	Val	Ile	Met	Asp	Gln	GΙΆ	
	770					775					780					
					•											
=							<b></b> -		c - +		aa+	cac	+++	att	cca	2400
tca	ctg	gca	cta	agt	gga	ggt	gca	gac	Cat.	gga	990	cac	-1	900	209	
Ser	Leu	Ala	Leu	Ser	Gly	Gly	Ala	Asp	His	Gly	Gly	His	Pne	vai	Pro	
785					790					795					800	
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cca	CCL	gaa	555	gee	aac	cac		990		0	0	9	Com	כיכ	cor	
Pro	Pro	Glų	Phe	Val	Asn	His	Thr	GIA	•	ser	ser	GIU	ser		261	
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+	220	tos	gag	222	aat	.tta	tat	act	gat	tta	qqa	tta	gqt	act	gct	2496
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Ser	Asn	Ser	Glu	ràa	GIA	ьeu	Tyr		GIĀ	nea	GIA	neu		T111	774	
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Gly Asp Ile Thr Met Ile His Ser Glu Ile Leu Gln Tyr Ser Gln His 840 835 ttc caa gaa aga ggt tta cct cat ggt aat ggg ttt gct act att tta 2592 Phe Gln Glu Arg Gly Leu Pro His Gly Asn Gly Phe Ala Thr Ile Leu 855 cga act gtc gat agt gtt aac agt act aat gat ggg tta att tat act Arq Thr Val Asp Ser Val Asn Ser Thr Asn Asp Gly Leu Ile Tyr Thr 870 865 tat gat agt aaa tat ttg cag tca cca aga gta agt gat ttg aaa gat Tyr Asp Ser Lys Tyr Leu Gln Ser Pro Arg Val Ser Asp Leu Lys Asp 890 885 gcc atg tca aca cat agg ggt ata agg tta tct aaa cca aat ttt ggt 2736 Ala Met Ser Thr His Arg Gly Ile Arg Leu Ser Lys Pro Asn Phe Gly 905 900 ggt gcc aat gga act gct aat atg acg gat tct gct tct aca tcc aat 2784 Gly Ala Asn Gly Thr Ala Asn Met Thr Asp Ser Ala Ser Thr Ser Asn 915 920 gga tot gtg ttg aat aaa aat atg caa act aca gat gtt gat tca att 2832 Gly Ser Val Leu Asn Lys Asn Met Gln Thr Thr Asp Val Asp Ser Ile 935 930 tta agt ggt ctt gaa agt gaa gac gaa gct gcg ttt gtt gtt taa 2877 Leu Ser Gly Leu Glu Ser Glu Asp Glu Ala Ala Phe Val Val 950 945 <210> 2 <211> 958 <212> PRT <213> Candida albicans <400> 2 Met Asn Leu Phe Gln His Lys His Gln Lys Leu Ile Leu Gln Cys Tyr 10 Pro Ala Gly Lys Ala Val Asp Lys Lys Pro Asn Ser Ser Glu Leu Ser 25

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Tyr Leu Leu Tyr Tyr Ala Ser Thr Arg Arg Val Lys Leu Glu Lys Val

35 -

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Thr	Gly 130	Asp	Lys	Glu	Phe	Ile 135	Lys	Ile	Phe	Thr	Glu 140	Val	Phe	Gln	Thr
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Arg	Leu 210	Lys	Ser	Asn	Leu	His 215	Val	Glu	Asp	Asp	Gly 220	Lys	Arg	Leu	Ser
Arg 225	Ala	His	Leu	Gln	Lys 230	Ser	His	Ser	Lys	Ile 235	Ala	Gln	Gln	Ile	Asp 240
Asp	Asp	Phe	Thr	Asn 245	Asp	Ser	Leu	Thr	Leu 250	Thr	Asp	Ile	Thr	Glu 255	Lys
Ala	Phe	Ser	Ser 260	Met	Lys	Ser	Phe	Phe 265	Asn	Thr	Asn	Ala	Ala 270	Ser	Gln
Ile	Ser	Glu 275	Val	Thr	Arg		Val .280	Val	Gln	His	Asn	11e 285	Leu	Asn	Gly
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Gly	Leu	Leu	Ala	Pro 485	Gln	Gly	Asp		Asp 490		Asn	Arg	Lys	Met 495	Ile
Ile	Ser	Thr	Thr 500		Leu	Ile	Asn	Ile 505		Ala	Arg	Tyr	Leu 510	Lys	Val
Phe	Asp	Glu 515		Leu	Asn	Asn	Glu 520		Ala	Val	Gly	Asn 525	Ser	Lys	Гуз
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Ala 545		Glu	Gly	Val	Asn 550		Ser	: Asp	Asp	555		Asn	Gly	Lys	Asp 560
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565 570 575

Ile Ser His Phe Leu Met Tyr Ile Asp Lys Phe Phe Glu Asn Tyr Asp 580 585 590

Ser Pro Asn Thr Gin Ser Val Leu Leu Leu Val Thr Val Leu Lys Asp 595 600 605

Met Met Asn Ile Leu Gly Leu Asn Phe Leu Ser Asn Phe Ile Pro Phe 610 615 620

Phe His His Trp Val Met Lys Val Asn Arg Ala Ser Asn Phe Thr Gln 625 630 635 640

Arg Gln Lys Phe Lys Asp Thr Phe Ala His Ile Ile Leu Tyr Tyr Met 645 650 655

Leu Lys Asp Leu Asp Glu Gln Tyr Ser His Asp Leu Gln Asn Tyr Cys
660 665 670

Lys Ser Ser Lys Leu Phe Lys Gln Ile Leu Asp Ala Val Glu Tyr Arg 675 680 685

Lys Met Gln Lys Phe Trp Val His Gly Ile Asp Pro Ser Pro Ser Asp 690 695 700

Leu Glu Asn Thr Lys Gly Asp Arg Thr Ile Pro Thr Asp Ala Asn Gly
705 710 715 720

Asn Tyr Ile Ala Ile Arg Ile Lys Pro Glu Asn Ile Glu Glu Phe Ala . 725 730 735

Cys Gly Asn Asn Phe Leu Ile Val Trp Leu His Pro Gln Lys Gln Leu 740 745 750

Leu Thr Glu Ile Glu Lys Ser Gln Val Ser Thr His Met Ser Thr Phe
755 760 765

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770 775 780

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## SEQUENCE LISTING

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<110> Hoechst Marion Roussel
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<120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES

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<170> PatentIn Ver. 2.1

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<223> Description of Artificial Sequence:Homologous fragment to Sc YMR212c

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#### SEQUENCE LISTING

<110> Hoechst Marion Roussel <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES <130> SEQID13 <140> <141> <160> 2 <170> PatentIn Ver. 2.1 <210> 1 <211> 3771 <212> DNA <213> Candida albicans <220> <221> CDS <222> (1)..(3771) <220> <221> gene <222> (1)..(3771) <223> Gene CaDR325 <400> 1 atg gat ata cca cca aaa cca act ctt aag gca att aag aaa ttt aga 48 Met Asp Ile Pro Pro Lys Pro Thr Leu Lys Ala Ile Lys Lys Phe Arg 15 5 1 act ttg gat gaa ata aaa tat gcc atg aaa cat gtt ttc caa gat gct Thr Leu Asp Glu Ile Lys Tyr Ala Met Lys His Val Phe Gln Asp Ala 25 20 caa tta ggt tta gca gga cat aga aaa tta gtg gta att ttg aaa aat 144 Gln Leu Gly Leu Ala Gly His Arg Lys Leu Val Val Ile Leu Lys Asn 40 35 gta ttt aaa aaa gcc att gaa tta aat caa att aat ttc ttt gcc atg 192 Val Phe Lys Lys Ala Ile Glu Leu Asn Gln Ile Asn Phe Phe Ala Met 55 50 tgt ttt act aaa ttg tta tct aaa gta tta cct ttg aaa aga gga gtt

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cca Pro 145	att Ile	tcg Ser	gaa Glu	ttc Phe	ata Ile 150	tca Ser	tat Tyr	ttg Leu	ata Ile	aaa Lys 155	tat Tyr	tta Leu	ttg Leu	agt Ser	999 Gly 160	480
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Thr	Phe	Tyr	Gln	Tyr	Cys	Asn	Glu	Asn	Gln	Leu	His	Ala	Leu	Met	Asp	
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Ala	Asn	Phe	Pro	Glu	Leu	Leu	Asp	Leu	Ser	Ile	Thr	Leu			Tyr	
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Lev	. Ser	Val	Arg	Leu	Lys	Thr			Glu	Asn	Glu			. vaı	гÀг	
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											<b></b>			C 2 2	ato	1392
aca	tgg	gaa	act	tat	aat	gcc	aaç	att	gac	gaa	tta	gat	. gat	. Caa	ata	2374

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705					/10										. • •	
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785			•		790					795					800	
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GIu	vai	ьys		тте	Gly	mec	пÀг		116	val	vsħ	116		nia		
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Glu					gca Ala					Trp						3072
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Thr Asp Gly	Lys Asn Lys	Leu Glu Leu	•	Lys Pro Ile Thr	
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<212> PRT

<213> Candida albicans

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Val Phe Lys Lys Ala Ile Glu Leu Asn Gln Ile Asn Phe Phe Ala Met 50 55 60

Cys Phe Thr Lys Leu Leu Ser Lys Val Leu Pro Leu Lys Arg Gly Val 65 70 75 80

Leu Ala Gly Asp Arg Ile Val Lys Phe Cys Tyr Leu Phe Val Asn Gly 85 90 95

Leu Val Lys Asp Ala Asn Glu Glu Lys Arg Ser Lys Glu Glu Glu Lys
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Glu Glu Lys Asp Lys Asp Glu Asp Lys Asp Thr Asn Glu Ser Asp Lys
115 120 125

Asn Glu Glu Asp Gln Glu Asp Gln Glu Gly Glu Gly Asp Gln Glu Thr
130 135 140

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Ala	Tyr	Leu	Val 180	Glu	Phe	Leu	Thr	Glu 185`		His	Glu	Asn	Asn 190	Thr	Leu
Glu	Ala	Leu 195	Tyr	Thr	Leu	Leu	Ser 200	Asn	Arg	Leu	Gln	Asp 205	Lys	Glu	Leu
Ser	Ile 210	Arg	Ile	Gln	Ala	Val 215	Val	Ala	Leu	Ser	His 220	Phe	Gln	Leu	Phe
Glu 225	Phe	Ser	Ile	Glu	Gly 230	Asp	Thr	Gly	Glu	Phe 235	Glu	Asp	Glu	Leu	11e 240
Ser	Ser	Asn	Gln	Ile 245	Gln	Asn	Lys	Leu	Ile 250	Asn	Ser	Ile	Gln	Asn 255	Asp
Asp	Ser	Pro	Glu 260	Val	Arg	Arg	Ala	Ala 265	Leu	Met	Asn	Leu	Val 270	Lys	Thr
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Gly	Leu	Asn	Asp	Arg 325	Asp	Glu	Thr	Val	Lys 330	Ala	Ala	Ala	Thr	Lys 335	Met
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CT/EP99/07376

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Leu	Ser	Val 435	Arg	Leu	Lys	Thr	Ile 440	Asn	Glu	Asn	Glu	Asn 445	Leu	Val	Lys
Thr	Trp 450	Glu	Thr	Tyr	Asn	Ala 455	Lys	Ile	Asp	Glu	Leu 460	Asp	Asp	Gln	Ile
Phe 465	Ser	Leu	Glu	Asn	Gln 470	Ile	Ser	Arg	Ile	Asn 475	Thr	Asp	Ala	Asp	Asn 480
Phe	Arg	Lys	Ser	Leu 485	Ser	Asn	Ile	Glu	Glu 490	Asp	Ile	Ile	Glu	Ile 495	Asn
Ile	Ala	Lys	Asp 500	Leu	Phe	Lys	Lys	Arg 505	Ile	Lys	Gln	Leu	Lys 510	Asn	Asn
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Asp 545	Ile	Asn	Lys	Asn	Leu 550	Asp	Glu	Ile	Glu	His 555	His	Pro	Glu	Asp	Ile 560
Thr	Ala	Lys	Leu	Glu 565	Glu	Leu	Gln	Thr	Lys 570	Tyr	Asp	Ser	Cys	Ile 575	Arg
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Asp Tyr Gly Asp Glu Met Ala Arg Arg Lys Leu Leu His Ile Ile Arg

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Leu Arg Val Leu Arg Ala Leu Ser Ile Asn Glu Lys Asp Phe Val Ser

630

645

625

635

670

1et	Ala	Val 675	Glu	Ile	Ile	Thr	Asp 680	Ile	Arg	Asp	Ser	Arg 685	Asp	Asp	Glu
Slu	Phe 690	His	Ser	Ala	Ala	Ala 695	Thr	Phe	Asp	Asp	Asp 700	Asp	Asp	Asp	Ile
Leu 705	Gly	Asn	Gly	Asp	Asp 710	Glu	Ser	Gln	Gln	Ser 715	Ser	Ser	Leu	Ser	Ala 720
/al	Thr	Lys	Lys	Arg 725	Arg	Ile	Glu	Pro	Asp 730	Met	Pro	Pro	Asp	Asp 735	Ile
/al	Leu	Arg	Cys 740	Leu	Thr	Met	Thr	Gln 745	Tyr	Val	Leu	Glu	Val 750	Ile	Thr
His	Ser	Leu 755	Asp	Asp	His	Leu	Ser 760	Leu	Ser	Ser	Ile	Tyr 765	Ser	Gly	Ile
Val	Asn 770	Tyr	Ala	Ile	Gln	Asn 775	Glu	Ser	Lys	Lys	Lys 780	Leu	Tyr	Leu	Ala
31y 785	Leu	Thr	Cys	Leu	Gly 790	Leu	Tyr	Ser	Leu	Ile 795	Asp	Ser	Lys	Ile	Ala 800
Arg	Ile		Thr	Thr 805	Thr	Leu	Leu	Leu	Ala 810	Met	Arg	Ser	Asn	Gly 815	Glu
Glu	Val	Lys	Glu 820	Ile	Gly	Met	Lys	Ala 825	Ile	Val	Asp	Ile	Leu 830	Ala	Ile
Tyr	Gly	Met 835	Ser	Ile	Leu	Asp	Lys 840	Ser	Ser	Lys	Tyr	Lys 845	Tyr	Ser	Arg
Met 	Phe 850		Lys	Val	Leu	Asn 855		Phe	Asp	Ala	Pro 860	Lys	Leu	Gln	Сув
Ile 865	Val	Ala	Glu	Gly	Leu 870	Cys	Lys	Leu	Phe	Leu 875	Ala	Asp	Ile	Leu	Tyr 880
Lys	Thr	Asp	Lys	Arg 885	Ser	Leu	Phe	Gly	Asn 890	Ala	Ile	Gln	Gly	Gly 895	Gly
Gly	Gly	Gly	Gly 900	Gly	Asn	Asp	Asp	Pro 905	Thr	Thr	Thr	Asn	Asp 910	Asp	Glu
Thr	Glu	Glu 915		Thr	Asp	Arg	Glu 920	His	Glu	Lys	His	Leu 925	Phe	Glu	Ala

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Ile	Val	Leu	Ile	Tyr	Phe	Asn	Pro	Asn	Thr	Lys	Ser	Asn	Gln	Glu	Let
	930					935					940				
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	Gln	Ile	Leu	Ser		Cys	He	Pro	vai	955	·Ald	PHE	SEL	UIP	960
945					950					,,,					-
Asn	His	Gln	Ile	Asn	Leu	Ala	Ala	Val	Ser	Gly	Asp	Val	Ile	Tyr	Arg
				965					970					975	
											_	•		_,	_
Leu	Phe	Thr		Thr	Glu	Thr	Glu		Ser	Pro	Ser	Val	990	11e	Pro
			980					985					330		
Gln	Leu	Tle	Ser	Trp	Cvs	qsA	Pro	Arg	Asn	Leu	Val	Lys	Leu	Ser	Aşı
<b>01</b>	204	995			-,-		1000	Ī				1005			
Glu	Glu	Ile	Asn	Gln	Ala	Thr	Ser	His	Leu			Cys	Val	Tyr	Le
:	1010				1	1015					1020				
T a	Gln	บาไ	บาไ	Glu	Gln	Val	Asn	Δla	Ara	Asn	Val	Lys	Arq	Cys	Ile
ьец 025	GIII	vaı	vai		L030	Vai	Kup			1035		_,	•		104
023				_											
Ile	Asn	Asn	Leu	Asn	Lys	Phe	His	Ile	Thr	Glu	Glu	Leu	Glu	Ser	Ası
			1	1045					1050					1055	
	Leu	_,		•	-1-	<b>T</b>	N1 -	T 011	λen	בומ	Thr	Val	Glu	Leu	Ph
GIN	Leu		A1A 1060	Leu	116	пув		1065	vob	niu	****		1070		
		•	1000												
Thr	Asn	Asn	Glu	Asp	Asn	Pro	Asn	Phe	Ile	Leu	Asp	Lys	Pro	Thr	Ly
		1075					1080					1085			
									T1-	7	200	Two	Leu	Glu	т 1
	Asn	Phe	Asp	Thr		11e 1095	GIu	ser	11e		1100	Lys	пец	Giu	11
	1090				•	1093									
Ala	Gln	Lys	Arg	Glu	Glu	Asn	Glu	Leu	Ile	Lys	Ser	Gly	Thr	Asn	Se
105		-			1110					1115					112
														_	
Ile	Leu	His			Asp	Asp	Leu			Gly	Thr	Gly			Se
			. :	1125					1130					1135	
Gla	Ile	Ser	Ile	Lva	Ser	Glu	Thr	Lvs	Arg	Arg	Asp	Leu	Asp	Arg	Se
Ų I II			1140	-,, 5				-/- 1145			•		1150	_	
Leu	Gln	Val	Ser	Lys	Thr	Thr	Ser	Pro	Glu	Thr	Ser	Glu	Asn	Glu	As

Glu Glu Asp Asp Asn Glu Glu Glu Glu Glu Lys Lys Lys Ser Phe

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Thr Asp Gly Lys Asn Lys Leu Glu Leu Lys Ala Asp Lys Pro Ile Thr
185 1190 1195 1200

Phe Lys Ala Glu Asp Lys Arg Glu Gly Ser Val Glu Thr Asp His Gly 1205 1210 1215

Gln Glu Gln Val Leu Val Glu Ser Lys Lys Val Ile Asp Ser Asn Val 1220 1225 1230

Glu Asp Ser Leu Glu Asp Ile Asp Lys Phe Leu Glu Glu Ala Asp Asp 1235 1240 1245

Val Asp Tyr Gly Asp Ile Ser Met Asp 1250 1255

PCT/EP99/07376

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## SEQUENCE LISTING

- <110> Hoechst Marion Roussel
- <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES
- <130> SEQID14

<140>

<141>

<160> 1

<170> PatentIn Ver. 2.1

<210> 1

<211> 603

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Homologous
 fragment to Sc YDR325

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## SEQUENCE LISTING

- <110> Hoechst Marion Roussel
- <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES
- <130> SEQID15

<140>

<141>

<160> 1

<170> PatentIn Ver. 2.1

<210> 1

<211> 581

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Homologous fragment to Sc YDR325

#### <400> 1

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# SEQUENCE LISTING

## <110> Hoechst Marion Roussel

- <120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR SCRENING ANTIMYCOTIC SUBSTANCES USING SAID GENES
- <130> SEQID16

<140>

<141>

<160> 1

<170> PatentIn Ver. 2.1

<210> 1

<211> 662

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Homologous
 fragment to Sc YDR325

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#### SEQUENCE LISTING

<110> Hoechst Marion Roussel

<120> ESSENTIAL GENES FROM C. ALBICANS AND A METHOD FOR
 SCREENING SUBSTANCES USING SAID GENES

<130> SEQID17

<140>

<141>

<160> 1

<170> PatentIn Ver. 2.1

<210> 1

<211> 231

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Homologous fragment to Sc YOR110

<400> 1

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